Field of the Invention

This invention relates to proteins having serine/threonine kinase domains, corresponding nucleic acid molecules, and their use.

Background of the Invention

The transforming growth factor-B (TGF-B) superfamily consists of a family of structurally-related proteins, including three different mammalian isoforms of TGF-B (TGF-B1, B2 and B3), activins, inhibins, müllerian-inhibiting substance and bone morphogenic proteins (BMPs) (for reviews see Roberts and Sporn, (1990) Peptide Growth Factors and Their Receptors, Pt.1, Sporn and Roberts, eds. (Berlin: Springer - Verlag) pp 419-472; Moses et al (1990) Cell 63, 245-247). The proteins of the TGF-B superfamily have a wide variety of biological activities. TGF-B acts as a growth inhibitor for many cell types and appears to play a central role in the regulation of embryonic development, tissue regeneration, immuno-regulation, as well as in fibrosis and carcinogenesis (Roberts and Sporn (199) see above).

Activins and inhibins were originally identified as factors which regulate secretion of follicle-stimulating hormone secretion (Vale et al (1990) Peptide Growth Factors and Their Receptors, Pt.2, Sporn and Roberts, eds. (Berlin: Springer-Verlag) pp.211-248). Activins were also shown to induce the differentiation of haematopoietic progenitor cells (Murata et al (1988) Proc. Natl. Acad. Sci. USA 85, 2434 - 2438; Eto et al (1987) Biochem. Biophys. Res. Commun. 142, 1095-1103) and induce mesoderm formation in Xenopus embryos (Smith et al (1990) Nature 345, 729-731; van den Eijnden-Van Raaij et al (1990) Nature 345, 732-734).

EMPs or osteogenic proteins which induce the formation of bone and cartilage when implanted subcutaneously (Wozney et al (1988) Science 242, 1528-1534), facilitate neuronal

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differentiation (Paralkar et al (1992) J. Cell Biol. 119, 1721-1728) and induce monocyte chemotaxis (Cunningham et al (1992) Proc. Natl. Acad. Sci. USA 89, 11740-11744). Müllerian-inhibiting substance induces regression of the Müllerian duct in the male reproductive system (Cate et al (1986) Cell 45, 685-698), and a glial cell line-derived neurotrophic factor enhances survival of midbrain dopaminergic neurons (Lin et al (1993) Science 260, 1130-1132). The action of these growth factors is mediated through binding to specific cell surface receptors.

Within this family, TGF-B receptors have been most thoroughly characterized. By covalently cross-linking radio-labelled TGF-B to cell surface molecules followed by polyacrylamide gel electrophoresis of the affinity-labelled complexes, three distinct size classes of cell surface proteins (in most cases) have been identified, denoted receptor type I (53 kd), type II (75 kd), type III or betaglycan (a 300 kd proteoglycan with a 120 kd core protein) (for a review see Massague (1992) Cell 69 1067-1070) and more recently endoglin (a homodimer of two 95 kd subunits) (Cheifetz et al (1992) J. Biol. Chem. 267 19027-19030). Current evidence suggests that type I and type II receptors are directly involved in receptor signal transduction (Segarini et al (1989) Mol. Endo., 2, 261-272; Laiho et al (1991) J. Biol. Chem. 266, 9100-9112) and may form a heteromeric complex; the type II receptor is needed for the binding of TGP-B to the type I receptor and the type I receptor is needed for the signal transduction induced by the type II receptor (Wrana et al (1992) Cell, 71, 1003-1004). The type III receptor and endoglin may have more indirect roles, possibly by facilitating the binding of ligand to type II receptors (Wang et al (1991) Cell, 67 797-805; López-Casillas et al (1993) Cell, 73 1435-1444).

Binding analyses with activin A and BMP4 have led to the identification of two co-existing cross-linked affinity complexes of 50-60 kDa and 70-80 kDa on responsive cells

(Hino et al (1989) J. Biol. Chem. 264, 10309 - 10314; Mathews and Vale (1991), Cell 68, 775-785; Paralker et al (1991) Proc. Natl. Acad. Sci. USA 87, 8913-8917). By analogy with TGF-B receptors they are thought to be signalling receptors and have been named type I and type II receptors.

Among the type II receptors for the TGF-ß superfamily of proteins, the cDNA for the activin type II receptor (Act RII) was the first to be cloned (Mathews and Vale (1991) Cell 65, 973-982). The predicted structure of the receptor was shown to be a transmembrane protein with an intracellular serine/threonine kinase domain. The activin receptor is related to the C. elegans daf-I gene product, but the ligand is currently unknown (Georgi et al (1990) Cell 61, 635-645). Thereafter, another form of the activin type II receptor (activin type IIB receptor), of which there are different splicing variants (Mathews et al (1992), Science 225, 1702-1705; Attisano et al (1992) Cell 68, 97-108), and the TGF-ß type II receptor (TBRII) (Lin et al (1992) Cell 68, 775-785) were cloned, both of which have putative serine/threconine kinase domains.

Summary of the Invention

The present invention involves the discovery of related novel peptides, including peptides having the activity of those defined herein as SEQ ID Nos. 2, 4, 8, 10, 12, 14, 16 and 18. Their discovery is based on the realisation that receptor serine/threonine kinases form a new receptor family, which may include the type II receptors for other proteins in the TGF-8 superfamily. To ascertain whether there were other members of this family of receptors, a protocol was designed to clone ActRII/daf I related cDNAs. This approach made use of the polymerase chain reaction (PCR), using degenerate primers based upon the amino-acid sequence similarity between kinase domains of the mouse activin type II receptor and daf-I gene products.

This strategy resulted in the isolation of a new family of receptor kinases called Activin receptor like kinases (ALK's) 1-6. These cDNAs showed an overall 33-39% sequence similarity with ActRII and TGF-8 type II receptor and 40-92% sequence similarity towards each other in the kinase domains.

Soluble receptors according to the invention comprise at least predominantly the extracellular domain. These can be selected from the information provided herein, prepared in conventional manner, and used in any manner associated with the invention.

Antibodies to the peptides described herein may be raised in conventional manner. By selecting unique sequences of the peptides, antibodies having desired specificity can be obtained.

The antibodies may be monoclonal, prepared in known manner. In particular, monoclonal antibodies to the extracellular domain are of potential value in therapy.

products of the invention are useful in diagnostic methods, e.g. to determine the presence in a sample for an analyte binding therewith, such as in an antagonist assay. Conventional techniques, e.g. an enzyme-linked immunosorbent assay, may be used.

products of the invention having a specific receptor activity can be used in therapy, e.g. to modulate conditions associated with activin or TGF- β activity. Such conditions include fibrosis, e.g. liver cirrhosis and pulmonary fibrosis, cancer, rheumatoid arthritis and glomeronephritis.

30 Brief Description of the Drawings

Figure 1 shows the alignment of the serine/threonine (S/T) kinase domains (I-VIII) of related receptors from transmembrane proteins, including embodiments of the present invention. The nomenclature of the subdomains is accordingly to Hanks et al (1988).

Figures 2A to 2D shows the sequences and characteristics of the respective primers used in the

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initial PCR reactions. The nucleic acid sequences are also given as SEQ ID Nos. 19 to 22.

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Figure 3 is a comparison of the amino-acid sequences SEQ 10 NO:34

of human activin type II receptor (Act R-II), mouse activin type IIB receptor (Act R-IIB), human TGF-B type II receptor (SEQ 16 NO:10)

(TBR-II), human TGF-B type I receptor (ALK-5), human activin receptor type IA (ALK-2), and type IB (ALK-4), ALKS

SEQ 10 NO:22 and SEQ 10 NO:16

1 & 3, and mouse ALK-6.

Figure 4 shows, schematically, the structures for Daf-1, Act R-II, Act R-IIB, TBR-II, TBR-I/ALK-5, ALK's -1, -2 (Act RIA), -3, -4 (Act RIB) & -6.

elio 3/C

Figure 5 shows the sequence alignment of the cysteine(Position 30-100 t Sequence alignment of the cysteine(Position 30-100 t Sequence)

Tich domains of the ALKS, TBR-II, Act R-II, Act R-IIB and on 41-443-6-500 to do and the cysteine of Sequence alignment of the cysteine
(Position 30-150 of Sequence)

daf-1, receptors.

Figure 6 is a comparison of kinase domains of serine/threonine kinases, showing the percentage amino-acid identity of the kinase domains.

Figure 7 shows the pairwise alignment relationship between the kinase domains of the receptor serine/threonine kinases. The dendrogram was generated using the Jotun-Hein alignment program (Hein (1990) Meth. Enzymol. 183, 626-645).

Brief Description of the Sequence Listings

Sequences 1 and 2 are the nucleotide and deduced amino-acid sequences of cDNA for hALK-1 (clone HP57).

Sequences 3 and 4 are the nucleotide and deduced amino-acid sequences of cDNA for hALK-2 (clone HP53).

Sequences 5 and 6 are the nucleotide and deduced amino-acid sequences of cDNA for hALK-3 (clone ONF5).

Sequences 7 and 8 the nucleotide and deduced aminoacid sequences of cDNA for hALK-4 (clone 11H8), complemented with PCR product encoding extracellular domain.

Sequences 9 and 10 are the nucleotide and deduced amino-acid sequences of cDNA for hALK-5 (clone EMBLA).

Sequences 11 and 12 are the nucleotide and deduced amino-acid sequences of cDNA for mALK-1 (clone AM6).

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Sequences 13 and 14 are the nucleotide and deduced amino-acid sequences of cDNA for mALK-3 (clones ME-7 and ME-D).

Sequences 15 and 16 are the nucleotide and deduced amino-acid sequences of cDNA for mALK-4 (clone 8a1).

Sequences 17 and 18 are the nucleotide and deduced amino-acid sequences of cDNA for mALK-6 (clone ME-6).

Sequence 19 (B1-S) is a sense primer, extracellular domain, cysteine-rich region, BamHI site at 5' end, 28-mer, 64-fold degeneracy.

Sequence 20 (B3-S) is a sense primer, kinase domain II. BamHI site at 5' end, 25-mer, 162-fold degeneracy.

Sequence 21 (B7-S) is a sense primer, kinase domain VIB, S/T kinase specific residues, BamHI site at 5' end, 24-mer, 288-fold degeneracy.

Sequence 22 (E8-AS) is an anti-sense primer, kinase domain, S/T kinase-specific residues EcoRI site at 5' end, 20-mer, 18-fold degeneracy.

Sequence 23 is an oligonucleotide probe.

Sequence 24 is a 5' primer.

Sequence 25 is a 3' primer.

Sequence 26 is a consensus sequence in Subdomain I.

Sequences 27 and 28 are novel sequence motifs in Subdomain VIB.

25 Sequence 29 is a novel sequence motif in Subdomain VIII.

Description of the Invention

As described in more detail below, nucleic acid sequences have been isolated, coding for a new sub-family of serine/threonine receptor kinases. The term nucleic acid molecules as used herein refers to any sequence which codes for the murine, human or mammalian form, amino-acid sequences of which are presented herein. It is understood that the well known phenomenon of codon degeneracy provides for a great deal of sequence variation and all such varieties are included within the scope of this invention.

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The nucleic acid sequences described herein may be used to clone the respective genomic DNA sequences in order to study the genes' structure and regulation. The murine and human cDNA or genomic sequences can also be used to isolate the homologous genes from other mammalian species. The mammalian DNA sequences can be used to study the receptors' functions in various in vitro and in vivo model systems.

As exemplified below for ALK-5 cDNA, it is also recognised that, given the sequence information provided herein, the artisan could easily combine the molecules with a pertinent promoter in a vector, so as to produce a cloning vehicle for expression of the molecule. promoter and coding molecule must be operably linked via well-recognized any of the and easily-practised methodologies for so doing. The resulting vectors, as well as the isolated nucleic acid molecules themselves, may be used to transform prokaryotic cells (e.g. E. coli), or transfect eukaryotes such as yeast (S. cerevisiae), PAE, COS or CHO cell lines. Other appropriate expression systems will also be apparent to the skilled artisan.

Several methods may be used to isolate the ligands for the ALKs. As shown for ALK-5 cDNA, cDNA clones encoding the active open reading frames can be subcloned into expression vectors and transfected into eukaryotic cells, for example COS cells. The transfected cells which can express the receptor can be subjected to binding assays for radioactively-labelled members of the TGF-B superfamily (TGF-B, activins, inhibins, bone morphogenic proteins and müllerian-inhibiting substances), as it may be expected that the receptors will bind members of the TGP-B superfamily. Various biochemical or cell-based assays can be designed to identify the ligands, in tissue extracts or conditioned media, for receptors in which a ligand is not known. Antibodies raised to the receptors may also be used to identify the ligands, using the immunoprecipitation of the cross-linked complexes. Alternatively, purified

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receptor could be used to isolate the ligands using an affinity-based approach. The determination of the expression patterns of the receptors may also aid in the isolation of the ligand. These studies may be carried out using ALK DNA or RNA sequences as probes to perform in situ hybridisation studies.

The use of various model systems or structural studies should enable the rational development of specific agonists and antagonists useful in regulating receptor function. It may be envisaged that these can be peptides, mutated ligands, antibodies or other molecules able to interact with the receptors.

The foregoing provides examples of the invention Applicants intend to claim which includes, inter alia, isolated nucleic acid molecules coding for activin receptor-like kinases (ALKs), as defined herein. These include such sequences isolated from mammalian species such as mouse, human, rat, rabbit and monkey.

The following description relates to specific embodiments. It will be understood that the specification and examples are illustrative but not limitative of the present invention and that other embodiments within the spirit and scope of the invention will suggest themselves to those skilled in the art.

25 Preparation of mRNA and Construction of a cDNA Library

For construction of a cDNA library, poly (A) RNA was isolated from a human erythroleukemia cell line (HEL 92.1.7) obtained from the American Type Culture Collection (ATCC TIB 180). These cells were chosen as they have been shown to respond to both activin and TGP-B. Moreover leukaemic cells have proved to be rich sources for the cloning of novel receptor tyrosine kinases (Partanen et al (1990) Proc. Natl. Acad. Sci. USA 87, 8913-8917 and (1992) Mol. Cell. Biol. 12, 1698-1707). (Total) RNA was prepared by the guanidinium isothiocyanate method (Chirgwin et al (1979) Biochemistry 18, 5294-5299). mRNA was selected using the poly-A or poly AT tract mRNA isolation kit

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(Promega, Madison, Wisconsin, U.S.A.) as described by the manufacturers, or purified through an oligo (dT)-cellulose column as described by Aviv and Leder (1972) Proc. Natl. Acad. Sci. USA 69, 1408-1412. The isolated mRNA was used for the synthesis of random primed (Amersham) cDNA, that was used to make a lgt10 library with 1x105 independent CDNA clones using the Riboclone cDNA synthesis system (Promega) and Agt10 in vitro packaging kit (Amersham) according to the manufacturers' procedures. An amplified oligo (dT) primed human placenta 12APII cDNA library of 5x10⁵ independent clones was used. Poly (A) RNA isolated from AG1518 human foreskin fibroblasts was used to prepare a primary random primed AZAPII cDNA library of 1.5x106 independent clones using the RiboClone cDNA synthesis system and Gigapack Gold II packaging extract (Stratagene). In addition, a primary oligo (dT) primed human foreskin fibroblast Agt10 cDNA library (Claesson-Welsh et al (1989) Proc. Natl. Acad. Sci. USA. 86 4917-4912) was prepared. An amplified oligo (dT) primed HEL cell lgt11 cDNA library of 1.5 X 106 independent clones (Poncz et al (1987) Blood 69 219-223) was used. A twelve-day mouse embryo AEXTOX CDNA library was obtained from Novagen (Madison, Wisconsin, U.S.A.); a mouse placenta AZAPII cDNA library was also used.

25 Generation of cDNA Probes by PCR

For the generation of cDNA probes by PCR (Lee et al (1988) Science 239, 1288-1291) degenerate PCR primers were constructed based upon the amino-acid sequence similarity between the mouse activin type II receptor (Mathews and Vale (1991) Cell 65, 973-982) and daf-1 (George et al (1990) Cell 61, 635-645) in the kinase domains II and VIII. Figure 1 shows the aligned serine/threonine kinase domains (I-VIII), of four related receptors of the TGF-8 superfamily, i.e. hT&R-II, mActR-IIB, mActR-II and the daf-1 gene product, using the nomenclature of the subdomains according to Hanks et al (1988) Science 241, 45-52.

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Several considerations were applied in the design of the PCR primers. The sequences were taken from regions of homology between the activin type II receptor and the daf-1 gene product, with particular emphasis on residues that confer serine/threonine specificity (see Table 2) and on residues that are shared by transmembrane kinase proteins and not by cytoplasmic kinases. The primers were designed so that each primer of a PCR set had an approximately similar GC composition, and so that self complementarity and complementarity between the 3' ends of the primer sets were avoided. Degeneracy of the primers was kept as low as possible, in particular avoiding serine, leucine and arginine residues (6 possible codons), and human codon Degeneracy was particularly preference was applied. avoided at the 3' end as, unlike the 5' end, where mismatches are tolerated, mismatches at the 3' end dramatically reduce the efficiency of PCR.

In order to facilitate directional subcloning, restriction enzyme sites were included at the 5' end of the primers, with a GC clamp, which permits efficient restriction enzyme digestion. The primers utilised are shown in Figure 2. Oligonucleotides were synthesized using Gene assembler plus (Pharmacia - LKB) according to the manufacturers instructions.

The mRNA prepared from HEL cells as described above was reverse-transcribed into cDNA in the presence of 50 mM Tris-HCl, pH 8.3, 8 mM MgCl₂, 30 mM KCl, 10 mM dithiothreitol, 2mM nucleotide triphosphates, excess oligo (dT) primers and 34 units of AMV reverse transcriptase at 42°C for 2 hours in 40 µl of reaction volume. Amplification by PCR was carried out with a 7.5% aliquot (3 µl) of the reverse-transcribed mRNA, in the presence of 10 mM Tris-HCl, pH 8.3, 50 mM KCl, 1.5 M MgCl₂, 0.01% gelatin, 0.2 mM nucleotide triphosphates, 1 µM of both sense and antisense primers and 2.5 units of Taq polymerase (Perkin Elmer Cetus) in 100 µl reaction volume. Amplifications were performed on a thermal cycler (Perkin Elmer Cetus)

using the following program: first 5 thermal cycles with denaturation for 1 minute at 94°C, annealing for 1 minute at 50°C, a 2 minute ramp to 55°C and elongation for 1 minute at 72°C, followed by 20 cycles of 1 minute at 94°C, 30 seconds at 55°C and 1 minute at 72°C. A second round of PCR was performed with 3 μ l of the first reaction as a template. This involved 25 thermal cycles, each composed of 94°C (1 min), 55°C (0.5 min), 72°C (1 min).

General procedures such as purification of nucleic acids, restriction enzyme digestion, gel electrophoresis, transfer of nucleic acid to solid supports and subcloning were performed essentially according to established procedures as described by Sambrook et al, (1989), Molecular cloning: A Laboratory Manual, 2nd Ed. Cold Spring Harbor Laboratory (Cold Spring Harbor, New York, USA).

Samples of the PCR products were digested with BamHI and EcoRI and subsequently fractionated by low melting point agarose gel electrophoresis. Bands corresponding to the approximate expected sizes, (see Table 1: ≈460 bp for primer pair B3-S and E8-AS and ≈ 140 bp for primer pair B7-S and E8-AS) were excised from the gel and the DNA was purified. Subsequently, these fragments were ligated into pUC19 (Yanisch-Perron et al (1985) Gene 33, 103-119), which had been previously linearised with BamHI and EcoR1 and transformed into E. coli strain DH5α using standard protocols (Sambrook et al, supra). Individual clones were sequenced using standard double-stranded sequencing techniques and the dideoxynucleotide chain termination method as described by Sanger et al (1977) Proc. Natl. Acad. Sci. USA 74, 5463-5467, and T7 DNA polymerase.

Employing Reverse Transcriptase PCR on HEL mRNA with the primer pair B3-S and E8-AS, three PCR products were obtained, termed 11.1, 11.2 and 11.3, that corresponded to novel genes. Using the primer pair B7-S and E8-AS, an additional novel PCR product was obtained termed 5.2.

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TABLE 1

KAME OF PCR PRODUCT	PRIMERS	INSERT SISE (bp)	SISE OF DICA FRACESTIN BACTRII/ BISRII CLONES (bp)	SEQUENCE IDENTITY WITE SEQUENCE EACTRII/ETERII (%)	SEQUENCE IDENTITY BETWEEN BACTRII and TER-II (%)
11.1	B3-8/E8-AS	460	460	46/40	42
11.2	B3-8/E8-AS	460	460	49/44	47
11.3	B3-S/E8-AS	460	460	44/36	48
11.29	B3-5/28-AS	460	460	ND/100	ND
9.2	B1-S/E8-AS	800	795	100/ND	ND
5.2	B7-5/E8-AS	140	143	40/38	60

15 Isolation of cDNA Clones

The PCR products obtained were used to screen various cDNA libraries described <u>supra</u>. Labelling of the inserts of PCR products was performed using random priming method (Feinberg and Vogelstein (1983) Anal. Biochem, <u>132</u> 6-13) using the Megaprime DNA labelling system (Amersham). The oligonucleotide derived from the sequence of the PCR product 5.2 was labelled by phosphorylation with T4 polynucleotide kinase following standard protocols (Sambrook <u>et al</u>, <u>supra</u>). Hybridization and purification of positive bacteriophages were performed using standard molecular biological techniques.

The double-stranded DNA clones were all sequenced using the dideoxynucleotide chain-termination method as described by Sanger et al, supra, using T7 DNA polymerase (Pharmacia - LKB) or Sequenase (U.S. Biochemical Corporation, Cleveland, Ohio, U.S.A.). Compressions of nucleotides were resolved using 7-deaza-GTP (U.S. Biochemical Corp.) DNA sequences were analyzed using the DNA STAR computer program (DNA STAR Ltd. U.K.). Analyses of the sequences obtained revealed the existence of six

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distinct putative receptor serine/threonine kinases which have been named ALK 1-6.

To clone cDNA for ALK-1 the oligo (dT) primed human placenta cDNA library was screened with a radiolabelled insert derived from the PCR product 11.3; based upon their restriction enzyme digestion patterns, three different types of clones with approximate insert sizes. of 1.7 kb, 2 kb & 3.5 kb were identified. The 2 kb clone, named HP57, was chosen as representative of this class and subjected to complete sequencing. Sequence analysis of ALK-1 revealed a sequence of 1984 nucleotides including a poly-A tail (SEQ ID No. 1). The longest open reading frame encodes a protein of 503 amino-acids, with high sequence similarity to receptor serine/threonine kinases The first methionine codon, the putative below). translation start site, is at nucleotide 283-285 and is preceded by an in-frame stop codon. This first ATG is in a more favourable context for translation initiation (Kozak (1987) Nucl. Acids Res., 15, 8125-8148) than the second and third in-frame ATG at nucleotides 316-318 and 325-327. The putative initiation codon is preceded by a 5' untranslated sequence of 282 nucleotides that is GC-rich (80% GC), which is not uncommon for growth factor receptors (Kozak (1991) J. Cell Biol., 115, 887-903). The 3' untranslated sequence comprises 193 nucleotides and ends with a poly-A tail. No bona fide poly-A addition signal is found, but there is a sequence (AATACA), 17-22 nucleotides upstream of the poly-A tail, which may serve as a poly-A addition signal.

ALK-2 cDNA was cloned by screening an amplified oligo (dT) primed human placenta cDNA library with a radiolabelled insert derived from the PCR product 11.2. Two clones, termed HP53 and HP64, with insert sizes of 2.7 kb and 2.4 kb respectively, were identified and their sequences were determined. No sequence difference in the overlapping clones was found, suggesting they are both derived from transcripts of the same gene.

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Sequence analysis of cDNA clone HP53 (SEQ ID No. 3) revealed a sequence of 2719 nucleotides with a poly-A tail. The longest open reading frame encodes a protein of 509 The first ATG at nucleotides 104-106 agrees favourably with Rozak's consensus sequence with an A at position 3. This ATG is preceded in-frame by a stop codon. There are four ATG codons in close proximity further downstream, which agree with the Kozak's consensus sequence (Kozak, supra), but according to Kozak's scanning model the first ATG is predicted to be the translation start site. The 5' untranslated sequence is 103 nucleotides. untranslated sequence of 1089 nucleotides contains a polyadenylation signal located 9-14 nucleotides upstream from the poly-A tail. The cDNA clone HP64 lacks 498 nucleotides from the 5' end compared to HP53, but the sequence extended at the 3' end with 190 nucleotides and This suggests that different poly-A tail is absent. polyadenylation sites occur for ALK-2. In Northern blots, however, only one transcript was detected (see below).

The cDNA for human ALK-3 was cloned by initially screening an oligo (dT) primed human foreskin fibroblast CDNA library with an oligonucleotide (SEQ ID No. 23) derived from the PCR product 5.2. One positive cDNA clone with an insert size of 3 kb, termed ON11, was identified. However, upon partial sequencing, it appeared that this clone was incomplete; it encodes only part of the kinase domain and lacks the extracelluar domain. The most 5' sequence of ON11, a 540 nucleotide XbaI restriction encoding a truncated kinase domain. subsequently used to probe a random primed fibroblast cDNA library from which one cDNA clone with an insert size of 3 kb. termed ONF5, was isolated (SEQ ID No. 5). Sequence analysis of ONF5 revealed a sequence of 2932 nucleotides without a poly-A tail, suggesting that this clone was The longest open reading derived by internal priming. frame codes for a protein of 532 amino-acids. ATG codon which is compatible with Kozak's consensus

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sequence (Kozak, supra), is at 310-312 nucleotides and is preceded by an in-frame stop codon. The 5' and 3' untranslated sequences are 309 and 1027 nucleotides long, respectively.

ALK-4 cDNA was identified by screening a human oligo (dT) primed human erythroleukemia cDNA library with the radiolabelled insert of the PCR product 11.1 as a probe. One cDNA clone, termed 11H8, was identified with an insert size of 2 kb (SEQ ID No. 7). An open reading frame was found encoding a protein sequence of 383 amino-acids, encoding a truncated extracellular domain with high similarity to receptor serine/threonine kinases. The 3' untranslated sequence is 818 nucleotides and does not contain a poly-A tail, suggesting that the cDNA was internally primed. CDNA encoding the extracellular domain (nucleotides 1-366) was obtained from HEL cells by RT-PCR with 5' primer (SEQ ID No. 24) derived in part from sequence at translation start site of SKR-2 (a cDNA sequence deposited in GenBank data base, accesion number L10125, that is identical in part to ALK-4) and 3' primer (SEQ ID No. 25) derived from 11H8 cDNA clone.

ALK-5 was identified by screening the random primed HEL cell 1gt 10 cDNA library with the PCR product 11.1 as a probe. This yielded one positive clone termed EMBLA 25 (insert size of 5.3 kb with 2 internal EcoRI sites). Nucleotide sequencing revealed an open reading frame of 1509 bp, coding for 503 amino-acids. The open reading frame was flanked by a 5' untranslated sequence of 76 bp. and a 3' untranslated sequence of 3.7 kb which was not completely sequenced. The nucleotide and deduced aminoacid sequences of ALK-5 are shown in SEQ ID Nos. 9 and 10. In the 5' part of the open reading frame, only one ATG codon was found; this codon fulfils the rules of translation initiation (Kozak, supra). An in-frame stop codon was found at nucleotides (-54)-(-52) in the 5' untranslated region. The predicted ATG start codon is followed by a stretch of hydrophobic amino-acid residues

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which has characteristics of a cleavable signal sequence. Therefore, the first ATG codon is likely to be used as a translation initiation site. A preferred cleavage site for the signal peptidase, according to von Heijne (1986) Nucl. Acid. Res. 14, 4683-4690, is located between amino-acid residues 24 and 25. The calculated molecular mass of the primary translated product of the ALK-5 without signal sequence is 53,646 Da.

Screening of the mouse embryo LEX <u>Iox</u> cDNA library using PCR, product 11.1 as a probe yielded 20 positive clones. DNAs from the positive clones obtained from this digested library were with EcoRI and HindIII, electrophoretically separated on a 1.3% agarose gel and transferred to nitrocellulose filters according established procedures as described by Sambrook et al, supra. The filters were then hybridized with specific probes for human ALK-1 (nucleotide 288-670), ALK-2 (nucleotide 1-581), ALK-3 (nucleotide 79-824) or ALK-4 nucleotide 1178-1967). Such analyses revealed that a clone termed ME-7 hybridised with the human ALK-3 probe. However, nucleotide sequencing revealed that this clone was incomplete, and lacked the 5' part of the translated Screening the same cDNA library with a probe corresponding to the extracelluar domain of human ALK-3 (nucleotides 79-824) revealed the clone ME-D. This clone was isolated and the sequence was analyzed. Although this clone was incomplete in the 3' end of the translated region, ME-7 and ME-D overlapped and together covered the complete sequence of mouse ALK-3. The predicted amino-acid sequence of mouse ALK-3 is very similar to the human sequence; only 8 amino-acid residues differ (98% identity; see SEQ ID No. 14) and the calculated molecular mass of the primary translated product without the putative signal sequence is 57,447 Da.

of the clones obtained from the initial library screening with PCR product 11.1, four clones hybridized to the probe corresponding to the conserved kinase domain of

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ALK-4 but not to probes from more divergent parts of ALK-1 to -4. Analysis of these clones revealed that they have an identical sequence which differs from those of ALK-1 to -5 and was termed ALK-6. The longest clone ME6 with a 2.0 kb insert was completely sequenced yielding a 1952 bp fragment consisting of an open reading frame of 1506 bp (502 aminoacids), flanked by a 5' untranslated sequence of 186 bp, and a 3' untranslated sequence of 160 bp. The nucleotide and predicted amino-acid sequences of mouse ALK-6 are shown in SEQ ID Nos. 17 and 18. No polyadenylation signal was found in the 3' untranslated region of ME6, indicating that the cDNA was internally primed in the 3' end. Only one ATG codon was found in the 5' part of the open reading frame, which fulfils the rules for translation initiation (Kozak, supra), and was preceded by an in-frame stop codon at nucleotides 163-165. However, a typical hydrophobic leader sequence was not observed at the N terminus of the translated region. Since there is no ATG codon and putative hydrophobic leader sequence, this ATG codon is likely to be used as a translation initiation site. calculated molecular mass of the primary translated product with the putative signal sequence is 55,576 Da.

Mouse ALK-1 (clone AM6 with 1.9 kb insert) was obtained from the mouse placenta AZAPII cDNA library using human ALK-1 cDNA as a probe (see SEQ ID No. 11). Mouse ALK-4 (clone 8a1 with 2.3kb insert) was also obtained from this library using human ALK-4 cDNA library as a probe (SEQ ID No. 15).

To summarise, clones HP22, HP57, ONF1, ONF3, ONF4 and HP29 encode the same gene, ALK-1. Clone AM6 encodes mouse ALK-1. HP53, HP64 and HP84 encode the same gene, ALK-2. ONF5, ONF2 and ON11 encode the same gene ALK-3. ME-7 and ME-D encode the mouse counterpart of human ALK-3. 11H8 encodes a different gene ALK-4, whilst Sal encodes the mouse equivalent. EMBLA encodes ALK-5, and ME-6 encodes ALK-6.

The sequence alignment between the 6 ALK genes and TER-II, mActR-II and ActR-IIB is shown in Figure 3. These molecules have a similar domain structure; an N-terminal predicted hydrophobic signal sequence (von Heijne (1986) Nucl. Acids Res. 14: 4683-4690) is followed by a relatively small extracellular cysteine-rich ligand binding domain, a single hydrophobic transmembrane region (Kyte & Doolittle (1982) J. Mol. Biol. 157, 105-132) and a C-terminal intracellular portion, which consists almost entirely of a kinase domain (Figures 3 and 4).

The extracelluar domains of these receptors have cysteine-rich regions, but they show little sequence similarity; for example, less than 20% sequence identity is found between <u>Daf</u>-1, ActR-II, T&R-II and ALK-5. The ALKs appear to form a subfamily as they show higher sequence similarities (15-47% identity) in their extracellular domains. The extracellular domains of ALK-5 and ALK-4 have about 29% sequence identity. In addition, ALK-3 and ALK-6 share a high degree of sequence similarity in their extracellular domains (46% identity).

The positions of many of the cysteine residues in all receptors can be aligned, suggesting that the extracellular domains may adopt a similar structural configuration. See Figure 5 for ALKs-1,-2,-3 &- 5. Each of the ALKs (except ALK-6) has a potential N-linked glycosylation site, the position of which is conserved between ALK-1 and ALK-2, and between ALK-3, ALK-4 and ALK-5 (see Figure 4).

The sequence similarities in the kinase domains between daf-1, ActR-II, TBR-II and ALK-5 are approximately 40%, whereas the sequence similarity between the ALKs 1 to 6 is higher (between 59% and 90%; see Figure 6). Pairwise comparison using the Jutun-Hein sequence alignment program (Hein (1990) Meth, Enzymol., 183, 626-645), between all family members, identifies the ALKs as a separate subclass among serine/threonine kinases (Figure 7).

The catalytic domains of kinases can be divided into subdomains with stretches of conserved amino-acid

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residues. The key motifs are found in serine/threonine kinase receptors suggesting that they are functional kinases. The consensus seguence for the binding of ATP (Gly-X-Gly-X-X-Gly in subdomain I followed by a Lys residue further downstream in subdomain II) is found in all the ALKs.

The kinase domains of daf-1, ActR-II, and ALKs show approximately equal sequence similarity with tyrosine and serine/threonine protein kinases. However analysis of the amino-acid sequences in subdomains VI and VIII, which are the most useful to distinguish a specificity for phosphorylation of tyrosine residues versus serine/threonine residues (Hanks et al (1988) Science 241 42-52) indicates that these kinases are serine/threonine kinases; refer to Table 2.

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TABLE 2

	KINASE SUBDO		(AINS	
		VIB	AIII	
C.	Serine/threonine kinase consensus	DLKPEN SEQIDAD: 3 8	G (T/S) XX (Y/F) X _A	
5	Tyrosine kinase consensus	DLAARN FQ1600:39	XP(I/V) (K/R) W (T/M) SEQ WNO:44 A	
	Act R-II	DIKSKN Sequano;40	GTRRYM, SER 10 00:45	
	Act R-IIB	DFKSKN Seq 10H0:41	GTRRYM ^{SEQIONO'US}	
	TBR-II	DLKSSN Segidna45	GTARYM	
	ALK-I	DFKSRN Sea 10 Na 27	GTKRYM ^{SEQ} ID NO. 29	
10	ALK -2, -3, -4, -5, & -6	DLKSKN Seqidno:28	GTKRYM SEQ ID NO. 29	

The sequence motifs DLKSKN (Subdomain VIB) and GTKRYM (Subdomain VIII), that are found in most of the serine/threonine kinase receptors, agree well with the consensus sequences for all protein serine/threonine kinase receptors in these regions. In addition, these receptors, except for ALK-1, do not have a tyrosine residue surrounded by acidic residues between subdomains VII and VIII, which is common for tyrosine kinases. A unique characteristic of the members of the ALK serine/threonine kinase receptor family is the presence of two short inserts in the kinase

domain between subdomains VIA and VIB and between subdomains X and XI. In the intracellular domain, these regions, together with the juxtamembrane part and C-terminal tail, are the most divergent between family members (see Figures 3 and 4). Based on the sequence similarity with the type II receptors for TGF-B and activin, the C termini of the kinase domains of ALKs -1 to -6 are set at Ser-495, Ser-501, Ser-527, Gln-500, Gln-498 and Ser-497, respectively.

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The distribution of ALK-1, -2, -3, -4 was determined by Northern blot analysis. A Northern blot filter with mRNAs from different human tissues was obtained from Clontech (Palo Alto, C.A.). The filters were hybridized with 32P-labelled probes at 42°C overnight in 50% formaldehyde, 5 x standard saline citrate (SSC; 1xSSC is 50mM sodium citrate, pH 7.0, 150 mM NaCl), 0.1% SDS, 50 mM sodium phosphate, 5 x Denhardt's solution and 0.1 mg/ml order to minimize cross-In salmon sperm DNA. hybridization, probes were used that did not encode part of the kinase domains, but corresponded to the highly diverged sequences of either 5' untranslated and ligand-binding regions (probes for ALK-1, -2 and -3) or 3' untranslated sequences (probe for ALK-4). The probes were labelled by random priming using the Multiprime (or Mega-prime) DNA labelling system and $[\alpha^{-32}P]$ dCTP (Feinberg & Vogelstein (1983) Anal. Biochem. 132: 6-13). Unincorporated label was removed by Sephadex G-25 chromatography. Filters were washed at 65°C, twice for 30 minutes in 2.5 x SSC, 0.1% SDS and twice for 30 minutes in 0.3 x SSC, 0.1% SDS before being exposed to X-ray film. Stripping of blots was performed by incubation at 90-100°C in water for 20 minutes.

The ALK-5 mRNA size and distribution were determined by Northern blot analysis as above. An EcoR1 fragment of 980bp of the full length ALK-5 cDNA clone, corresponding to the C-terminal part of the kinase domain and 3'

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untranslated region (nucleotides 1259-2232 in SEQ ID No. 9) was used as a probe. The filter was washed twice in 0.5 x ssc. 0.1% SDS at 55°C for 15 minutes.

Using the probe for ALK-1, two transcripts of 2.2 and 4.9kb were detected. The ALK-1 expression level varied strongly between different tissues, high in placenta and lung, moderate in heart, muscle and kidney, and low (to not detectable) in brain, liver and pancreas. The relative ratios between the two transcripts were similar in most tissues; in kidney, however, there was relatively more of the 4.9 kb transcript. By reprobing the blot with a probe for ALK-2, one transcript of 4.0 kb was detected with a ubiquitous expression pattern. Expression was detected in every tissue investigated and was highest in placenta and skeletal muscle. Subsequently the blot was reprobed for One major transcript of 4.4 kb and a minor transcript of 7.9 kb were detected. Expression was high in skeletal muscle, in which also an additional minor transcript of 10 kb was observed. Moderate levels of ALK-3 mRNA were detected in heart, placenta, kidney and pancreas, and low (to not detectable) expression was found in brain, lung and liver. The relative ratios between the different transcripts were similar in the tested tissues, the 4.4 kb transcript being the predominant one, with the exception for brain where both transcripts were expressed at a similar level. Probing the blot with ALK-4 indicated the presence of a transcript with the estimated size of 5.2 kb and revealed an ubiquitous expression pattern. The results of Northern blot analysis using the probe for ALK-5 showed that a 5.5 kb transcript is expressed in all human tissues tested, being most abundant in placenta and least abundant in brain and heart.

The distribution of mRNA for mouse ALK-3 and -6 in various mouse tissues was also determined by Northern blot analysis. A multiple mouse tissue blot was obtained from Clontech, Palo Alto, California, U.S.A. The filter was hybridized as described above with probes for mouse ALK-3

and ALK-6. The <u>EcoRI-PstI</u> restriction fragment, corresponding to nucleotides 79-1100 of ALK-3, and the <u>SacI-HpaI</u> fragment, corresponding to nucleotides 57-720 of ALK-6, were used as probes. The filter was washed at 65°C twice for 30 minutes in 2.5 x SSC, 0.1% SDS and twice for 30 minutes with 0.3 x SSC, 0.1% SDS and then subjected to autoradiography.

Using the probe for mouse ALK-3, a 1.1 kb transcript was found only in spleen. By reprobing the blot with the ALK-6 specific probe, a transcript of 7.2 kb was found in brain and a weak signal was also seen in lung. No other signal was seen in the other tissues tested, i.e. heart, liver, skeletal muscle, kidney and testis.

All detected transcript sizes were different, and thus no cross-reaction between mRNAs for the different ALKs was observed when the specific probes were used. This suggests that the multiple transcripts of ALK-1 and ALK-3 are coded from the same gene. The mechanism for generation of the different transcripts is unknown at present; they may be formed by alternative mRNA splicing, differential polyadenylation, use of different promotors, or by a combination of these events. Differences in mRNA splicing in the regions coding for the extracellular domains may lead to the synthesis of receptors with different affinities for ligands, as was shown for mactr-IIB (Attisano et al (1992) Cell 68, 97-108) or to the production of soluble binding protein.

The above experiments describe the isolation of nucleic acid sequences coding for new family of human receptor kinases. The cDNA for ALK-5 was then used to determine the encoded protein size and binding properties. Properties of the ALKs cDNA Encoded Proteins

To study the properties of the proteins encoded by the different ALK cDNAs, the cDNA for each ALK was subcloned into a eukaryotic expression vector and transfected into various cell types and then subjected to immunoprecipitation using a rabbit antiserum raised against

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a synthetic peptide corresponding to part of the intracellular juxtamembrane region. This region is divergent in sequence between the various serine/threonine kinase receptors. The following amino-acid residues were

ALK-1 145-166

used:

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ALK-2 151-172

ALK-3 181-202

ALK-4 153-171

10 ALK-5 158-179

ALK-6 151-168

The rabbit antiserum against ALK-5 was designated VPN.

The peptides were synthesized with an Applied Biosystems 430A Peptide Synthesizer using t-butoxycarbonyl chemistry and purified by reversed-phase high performance liquid chromatography. The peptides were coupled to keyhole limpet haemocyanin (Calbiochem-Behring) using glutaraldehyde, as described by Guillick et al (1985) EMBO J. 4, 2869-2877. The coupled peptides were mixed with Freunds adjuvant and used to immunize rabbits.

Transient transfection of the ALK-5 cDNA

COS-1 cells (American Type Culture Collection) and the R mutant of MvILu cells (for references, see below) were cultured in Dulbecco's modified Eagle's medium containing 10% fetal bovine serum (FBS) and 100 units/ml penicillin and 50 µg lml streptomycin in 5% CO2 atmosphere at 37°C. The ALK-5 cDNA (nucleotides (-76) - 2232), which includes the complete coding region, was cloned in the pSV7d vector (Truett et al, (1985) DNA 4, 333-349), and used for transfection. Transfection into COS-1 cells was performed by the calcium phosphate precipitation method (Wigler et al (1979) Cell 16, 777-785). Briefly, cells were seeded into 6-well cell culture plates at a density of 5x105 cells/well, and transfected the following day with 10 μg of recombinant plasmid. After overnight incubation, cells were washed three times with a buffer containing 25 mM Tris-HCl, pH 7.4, 138 mM NaCl, 5 mM KCl, 0.7 mM CaCl, 0.5

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mM MgCl₂ and 0.6 mM Na₂HPO₄, and then incubated with Dulbecco's modified Eagle's medium containing FBS and antibiotics. Two days after transfection, the cells were metabolically labelled by incubating the cells for 6 hours in methionine and cysteine-free MCDB 104 medium with 150 μ Ci/ml of [35]-methionine and [35]-cysteine (in vivo labelling mix; Amersham). After labelling, the cells were washed with 150 mM NaCI, 25 mM Tris-HCl, pH 7.4, and then solubilized with a buffer containing 20mm Tris-HCl, pH 7.4, 150 mM NaCl, 10 mM EDTA, 1% Triton X-100, 1% deoxycholate, 1.5% Trasylol (Bayer) and 1 mM phenylmethylsulfonylfluoride (PMSF; Sigma). After 15 minutes on ice, the cell lysates were pelleted by centrifugation, and the supernatants were then incubated with 7 μ l of preimmune serum for 1.5 hours Samples were then given 50 μ l of protein λ at 4°C. Sepharose (Pharmacia-LKB) slurry (50% packed beads in 150 mM NaCl, 20 mM Tris-HCl, pH 7.4, 0.2% Triton X100) and incubated for 45 minutes at 4°C. The beads were spun down by centrifugation, and the supernatants (1 ml) were then incubated with either 7 µl of preimmune serum or the VPN antiserum for 1.5 hours at 4°C. For blocking, 10 μ g of peptide was added together with the antiserum. complexes were then given 50 µl of protein A-Sepharose (Pharmacia - LKB) slurry (50% packed beads in 150 mM NaCl, 20mM Tris-HCl, pH 7.4, 0.2% Triton X-100) and incubated for 45 minutes at 4°C. The beads were spun down and washed four times with a washing buffer (20 mM Tris-HCl, pH 7.4. 500 mM NaCI, 1% Triton X-100, 1% deoxycholate and 0.2% SDS), followed by one wash in distilled water. The immune complexes were eluted by boiling for 5 minutes in the SDSsample buffer (100 mM Tris-HCl, pH 8.8, 0.01% bromophenol blue, 36% glycerol, 4% SDS) in the presence of 10 mM DTT. and analyzed by SDS-gel electrophoresis using 7-15% polyacrylamide gels (Blobel and Dobberstein, (1975) J.Cell Biol. 67, 835-851). Gels were fixed, incubated with Amplify (Amersham) for 20 minutes, and subjected to A component of 53Da was seen. fluorography. This

component was not seen when preimmune serum was used, or when 10 µg blocking peptide was added together with the antiserum. Moreover, it was not detectable in samples derived from untransfected COS-1 cells using either preimmune serum or the antiserum.

Digestion with Endoglycosidase F

Samples immunoprecipitated with the VPN antisera obtained as described above were incubated with 0.5 U of endoglycosidase F (Boehringer Mannheim Biochemica) in a buffer containing 100 mM sodium phosphate, pH 6.1, 50 mM EDTA, 1% Triton X-100, 0.1% SDS and 1% B-mercaptoethanol at 37°C for 24 hours. Samples were eluted by boiling for 5 minutes in the SDS-sample buffer, and analyzed by SDS-polyacrylamide gel electrophoresis as described above. Hydrolysis of N-linked carbohydrates by endoglycosidase F shifted the 53 kDa band to 51 kDa. The extracelluar domain of ALK-5 contains one potential acceptor site for N-glycosylation and the size of the deglycosylated protein is close to the predicted size of the core protein.

20 Establishment of PAE Cell Lines Expressing ALK-5

In order to investigate whether the ALK-5 cDNA encodes a receptor for TGF-8, porcine aortic endothelial (PAE) cells were transfected with an expression vector containing the ALK-5 cDNA, and analyzed for the binding of ¹²⁵I-TGF-81.

PAE cells were cultured in Ham's F-12 medium supplemented with 10% FBS and antibiotics (Miyazono et al., (1988) J. Biol. Chem. 261, 6407-6415). The ALK-5 cDNA was cloned into the cytomegalovirus (CMV)-based expression vector pcDNA I/NEO (Invitrogen), and transfected into PAE cells by electroporation. After 48 hours, selection was initiated by adding Geneticin (G418 sulphate; Gibco - BRL) to the culture medium at a final concentration of 0.5 mg/ml (Westermark et al., (1990) Proc. Natl. Acad. Sci. USA 87, 128-132). Several clones were obtained, and after analysis by immunoprecipitation using the VPN antiserum, one clone denoted PAE/TBR-1 was chosen and further analyzed.

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Todination of TGF-81. Binding and Affinity Crosslinking

Recombinant human TGF-81 was iodinated using the chloramine T method according to Frolik et al., (1984) J. Biol. Chem. 259, 10995-11000. Cross-linking experiments were performed as previously described (Ichijo et al., (1990) Exp. Cell Res. 187, 263-269). Briefly, cells in 6well plates were washed with binding buffer (phosphatebuffered saline containing 0.9 mM CaCl,, 0.49 mM MgCl, and 1 mg/ml bovine serum albumin (BSA)), and incubated on ice in the same buffer with 125I-TGF-B1 in the presence or absence of excess unlabelled TGF-81 for 3 hours. were washed and cross-linking was done in the binding buffer without BSA together with 0.28 mM disuccinimidyl suberate (DSS; Pierce Chemical Co.) for 15 minutes on ice. The cells were harvested by the addition of 1 ml of detachment buffer (10 mM Tris-HCl, pH 7.4, 1 mM EDTA, 10% glycerol, 0.3 mM PMSF). The cells were pelleted by . centrifugation, then resuspended in 50 µl of solubilization buffer (125 mM NaCl, 10 mM Tris-HCl, pH 7.4, 1 mM EDTA, 1% Triton X-100, 0.3 mM PMSF, 1% Trasylol) and incubated for Cells were centrifuged again and 40 minutes on ice. supernatants were subjected to analysis by SDS-gel electrophoresis using 4-15% polyacrylamide gels, followed 125 I-TGF-B1 formed a 70 kDa crossby autoradiography. linked complex in the transfected PAE cells (PAE/TBR-Icells). The size of this complex was very similar to that of the TGF-B type I receptor complex observed at lower amounts in the untransfected cells. A concomitant increase of 94 kDa TGF-B type II receptor complex could also be observed in the PAE/TBR-I cells. Components of 150-190 kDa, which may represent crosslinked complexes between the type I and type II receptors, were also observed in the PAE/TBR-I cells.

In order to determine whether the cross-linked 70 kDa complex contained the protein encoded by the ALK-5 cDNA, the affinity cross-linking was followed by immunoprecipitation using the VPN antiserum. For this,

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cells in 25 cm2 flasks were used. The supernatants obtained after cross-linking were incubated with 7 μ l of preimmune serum or VPN antiserum in the presence or absence . of 10 μ g of peptide for 1.5h at 4°C. Immune complexes were then added to 50 μ l of protein λ -Sepharose slurry and incubated for 45 minutes at 4°C. The protein A-Sepharose beads were washed four times with the washing buffer, once with distilled water, and the samples were analyzed by SDSgel electrophoresis using 4-15% polyacrylamide gradient gels and autoradiography. A 70 kDa cross-linked complex was precipitated by the VPN antiserum in PAE/TBR-1 cells, and a weaker band of the same size was also seen in the untransfected cells, indicating that the untransfected PAE cells contained a low amount of endogenous ALK-5. The 70 kDa complex was not observed when preimmune serum was used, or when immune serum was blocked by 10 µg of peptide. Moreover, a coprecipitated 94 kDa component could also be observed in the PAE/TBR-I cells. The latter component is likely to represent a TGF-B type II receptor complex, since an antiserum, termed DRL, which was raised against a synthetic peptide from the C-terminal part of the TGF-B type II receptor, precipitated a 94 kDa TGP-B type II receptor complex, as well as a 70 kDa type I receptor complex from PAE/TBR-I cells.

The carbohydrate contents of ALK-5 and the TGF-8 type II receptor were characterized by deglycosylation using endoglycosidase F as described above and analyzed by SDS-polyacrylamide gel electrophoresis and autoradiography. The ALK-5 cross-linked complex shifted from 70 kDa to 66 kDa, whereas that of the type II receptor shifted from 94 kDa to 82 kDa. The observed larger shift of the type II receptor band compared with that of the ALK-5 band is consistent with the deglycosylation data of the type I and type II receptors on rat liver cells reported previously (Cheifetz et al (1988) J. Biol. Chem. 263, 16984-16991), and fits well with the fact that the porcine TGF-8 type II receptor has two N-glycosylation sites (Lin et al (1992)

Cell 68, 775-785), whereas ALK-5 has only one (see SEQ ID No. 9).

Binding of TGF-81 to the type I receptor is known to be abolished by transient treatment of the cells with dithiothreitol (DTT) (Cheifetz and Massague (1991) J. Biol. Chem. 266, 20767-20772; Wrana et al (1992) Cell 71, 1003-1014). When analyzed by affinity cross-linking, binding of 125 I-TGF-81 to ALK-5, but not to the type II receptor, was completely abolished by DTT treatment of PAE/TBR-1 cells. Affinity cross-linking followed by immunoprecipitation by the VPN antiserum showed that neither the ALK-5 nor the type II receptor complexes was precipitated after DTT treatment, indicating that the VPN antiserum reacts only The data show that the VPN antiserum with ALK-5. recognizes a TGF-B type I receptor, and that the type I and type II receptors form a heteromeric complex. 125 I-TGF-B1 Binding & Affinity Crosslinking of Transfected COS Cells

Transient expression plasmids of ALKs -1 to -6 and TBR-II were generated by subcloning into the pSV7d expression vector or into the pcDNA I expression vector (Invitrogen). Transient transfection of COS-1 cells and iodination of TGF-B1 were carried out as described above. Crosslinking and immunoprecipitation were performed as described for PAE cells above.

Transfection of cDNAs for ALKs into COS-1 cells did not show any appreciable binding of ¹²⁵I-TGF\$1, consistent with the observation that type I receptors do not bind TGF-8 in the absence of type II receptors. When the T\$R-II cDNA was co-transfected with cDNAs for the different ALKs, type I receptor-like complexes were seen, at different levels, in each case. COS-1 cells transfected with T\$R-II and ALK cDNAs were analyzed by affinity crosslinking followed by immunoprecipitation using the DRL antisera or specific antisera against ALKs. Each one of the ALKs bound ¹²⁵I-TGF-81 and was coimmunoprecipitated with the T\$R-II complex using the DRL antiserum. Comparison of the

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efficiency of the different ALRs to form heteromeric complexes with TBR-II, revealed that ALK-5 formed such complexes more efficiently than the other ALKs. The size of the crosslinked complex was larger for ALK-3 than for other ALKs, consistent with its slightly larger size. Expression of the ALK Protein in Different Cell Types

Two different approaches were used to elucidate which ALK's are physiological type I receptors for TGF-8.

Firstly, several cell lines were tested for the expression of the ALK proteins by cross-linking followed by immunoprecipitation using the specific antiseras against ALKs and the TGF-B type II receptor. The mink lung epithelial cell line, MvlLu, is widely used to provide target cells for TGF-B action and is well characterized regarding TGF-8 receptors (Laiho et al (1990) J. Biol. Chem. 265, 18518-18524; Laiho et al (1991) J. Biol. Chem. <u>266</u>, 9108-9112). Only the VPN antiserum efficiently precipitated both type I and type II TGF-8 receptors in the wild type Mv1Lu cells. The DRL antiserum also precipitated components with the same size as those precipitated by the VPN antiserum. A mutant cell line (R mutant) which lacks the TGF-B type I receptor and does not respond to TGF-B (Laiho et al, supra) was also investigated by cross-linking followed by immunoprecipitation. Consistent with the results obtained by Laiho et al (1990), supra the type III and type II TGF-B receptor complexes, but not the type I receptor complex, were observed by affinity crosslinking. Crosslinking followed by immunoprecipatition using the DRL antiserum revealed only the type II receptor complex. whereas neither the type I nor type II receptor complexes was seen using the VPN antiserum. When the cells were metabolically labelled and subjected to immunoprecipitation using the VPN antiserum, the 53 kDa ALK-5 protein was precipitated in both the wild-type and R mutant MviLu cells. These results suggest that the type I receptor expressed in the R mutant is ALK-5, which has lost the affinity for binding to TGF-B after mutation.

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The type I and type II TGF-B receptor complexes could be precipitated by the VPN and DRL antisera in other cell lines, including human foreskin fibroblasts (AG1518), human lung adenocarcinoma cells (A549), and human oral squamous cell carcinoma cells (HSC-2). Affinity cross-linking studies revealed multiple TGF-B type I receptor-like complexes of 70-77 kDa in these cells. These components were less efficiently competed by excess unlabelled TGF-81 in HSC-2 cells. Moreover, the type II receptor complex was low or not detectable in A549 and HSC-2 cells. linking followed by immunoprecipitation revealed that the VPN antiserum precipitated only the 70 kDa complex among the 70-77 kDa components. The DRL antiserum precipitated the 94 kDa type II receptor complex as well as the 70 kDa type I receptor complex in these cells, but not the putative type I receptor complexes of slightly larger These results suggest that multiple type I TGP-8 receptors may exist and that the 70 kDa complex containing ALK-5 forms a heteromeric complex with the TGF-8 type II receptor cloned by Lin et al (1992) Cell 68, 775-785, more efficiently that the other species. pheochromocytoma cells (PC12) which have been reported to have no TGF-B receptor complexes by affinity cross-linking (Hassagué et al (1990) Ann. N.Y. Acad. Sci. 593, 59-72), neither VPN nor DRL antisera precipitated the TGF-8 receptor complexes. The antisera against ALKs -1 to -4 and ALK6 did not efficiently immunoprecipitate the crosslinked receptor complexes in porcine aortic endothelial (PAE) cells or human foreskin fibroblasts.

Next, it was investigated whether ALKs could restore responsiveness to TGF-B in the R mutant of Mv1Lu cells, which lack the ligand-binding ability of the TGF-B type I receptor but have intact type II receptor. Wild-type Mv1Lu cells and mutant cells were transfected with ALK cDNA and were then assayed for the production of plasminogen activator inhibitor-1 (PAI-1) which is produced as a result of TGF-B receptor activation as described previously by

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Laiho et al (1991) Mol. Cell Biol. 11, 972-978. Briefly, cells were added with or without 10 ng/ml of TGF-81 for 2 serum-free MCDB 104 without methionine. Thereafter, cultures were labelled with [35s] methionine (40 μ Ci/ml) for 2 hours. The cells were removed by washing on ice once in PBS, twice in 10 mM Tris-HCl (pH 8.0), 0.5% sodium deoxycholate, 1 mM PMSF, twice in 2 mM Tris-HCl (pH 8.0), and once in PBS. Extracellular matrix proteins were extracted by scraping cells into the SDS-sample buffer containing DTT, and analyzed by SDS-gel electrophoresis followed by fluorography using Amplify. PAI-1 can be identified as a characteristic 45kDa band (Laiho et al (1991) Mol. Cell Biol. 11, 972-978). Wild-type MvILu cells responded to TGF-8 and produced PAI-1, whereas the R mutant clone did not, even after stimulation by TGF-81. Transient transfection of the ALR-5 cDNA into the R mutant clone led to the production of PAI-1 in response to the stimulation by TGF-81, indicating that the ALK-5 cDNA encodes a functional TGF-B type I receptor. In contrast, the R mutant cells that were transfected with other ALKs did not produce PAI-1 upon the addition of TGF-81.

Using similar approaches as those described above for the identification of TGF-B-binding ALKs, the ability of ALKs to bind activin in the presence of ActRII was examined. COS-1 cells were co-transfected as described above. Recombinant human activin A was indinated using the chloramine T method (Mathews and Vale (1991) Cell 65, 973-982). Transfected COS-1 cells were analysed for binding and crosslinking of ¹²⁵I-activin A in the presence or absence of excess unlabelled activin A. The crosslinked complexes were subjected to immunoprecipitation using DRL antisera or specific ALK antisera.

All ALKs appear to bind activin A in the presence of Act R-II. This is more clearly demonstrated by affinity cross-linking followed by immunoprecipitation. ALK-2 and ALK-4 bound 125 I-activin A and were coimmunoprecipitated

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with ActR-II. Other ALKs also bound 125 I-activin A but with a lower efficiency compared to ALK-2 and ALK-4.

In order to investigate whether ALKs are physiological activin type I receptors, activin responsive cells were examined for the expression of endogenous activin type I receptors. MylLu cells, as well as the R mutant, express both type I and type II receptors for activin, and the R mutant cells produce PAI-1 upon the addition of activin A. MylLu cells were labeled with ¹²⁵I-activin A, cross-linked and immunoprecipitated by the antisera against ActR-II or ALKs as described above.

The type I and type II receptor complexes in MvlLu cells were immunoprecipitated only by the antisera against ALK-2, ALK-4 and ActR-II. Similar results were obtained using the R mutant cells. PAE cells do not bind activin because of the lack of type II receptors for activin, and so cells were transfected with a chimeric receptor, to enable them to bind activin, as described herein. plasmid (chim A) containing the extracelluar domain and Cterminal tail of Act R-II (amino-acids -19 to 116 and 465 to 494, respectively (Mathews and Vale (1991) Cell, 65, 973-982)) and the kinase domain of TBR-II (amino-acids 160-543) (Lin et al (1992) Cell, 68, 775-785) was constructed and transfected into pcDNA/neo (Invitrogen). were stably transfected with the chim A plasmid by electroporation, and cells expressing the chim A protein were established as described previously. PAE/Chim A cells were then subjected to 12 I-activin A labelling crosslinking and immunoprecipitation as described above.

Similar to MvILu cells, activin type I receptor complexes in PAE/Chim A cells were immunoprecipitated by the ALK-2 and ALK-4 antisera. These results show that both ALK-2 and ALK-4 serve as high affinity type I receptors for activin A in these cells.

35 ALK-1, ALK-3 and ALK-6 bind TGF-81 and activin A in the presence of their respective type II receptors, but the

functional consequences of the binding of the ligands remains to be elucidated.

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The experiments described supra suggested further experiments. Specifically, it is known that TGF-S family members act as ligands in connection with specific type I and type II receptors, with resulting complexes interacting with members of the Smad family. See Heldin et al., Nature 390: 465-471 (1997), incorporated by reference. The Smad molecules are homologs of molecules found in Drosophila ("Mad"), and C. elegans (SMa), hence, the acronym These are involved in signal transduction pathways downstream of serine/threonine kinase receptors. See Massagué et al., Trends Cell Biol. 2: 187-192 (1997). The different members of the family have different signalling roles. Smad1, for example, as well as Smad 2 and 3, and perhaps Smad 5, become phosphorylated via specific type 1 serine/threonine kinase receptors, and act in pathway restricted fashion. For example, Xenopus Madl induces ventral mesoderm, in the presence of BMP. The human Smadl has been shown to have ventralizing activity. See Liu et al., Nature 381: 620-623 (1996); Kretzschmer et al., Genes Dev 11: 984-995 (1997). There is also some evidence that TGF-ß phosphorylates Smad1. Lechleider et al., J. Biol. Chem. 271: 17617-17620 (1996); Yingling et al., Proc. Natl. Acad. Sci. USA 93: 8940-8944 (1996). what was known regarding this complex signalling pathway, the role of ALK-1 was studied.

COS-7 cells, which do not express ALK-1, were transfected with cDNA encoding tagged ALK-1. The tag was hemagluttinin (hereafter "HA"), and a commercially available lipid containing transfecting agent was used. In parallel experiments, porcine aortic endothelial (PAE) cells were also used, because these cells express TGFS type II receptors, and ALK-5, but not ALK-1. Hence, PAE cells were either transfected, or not. Transfection protocols are given, supra.

The cells were then contacted with 125I labelled TGF-£1, and were then contacted with ALK-1 specific antisera, to ascertain

whether cross linking had occurred. See the experiments, supra, as well as ten Dijke et al., Science 264: 101-104 (1994), incorporated by reference. Antisera to ALK-5 were also used.

The results indicated that the ALK-1 antiserum immunoprecipitated complexes of the appropriate size from the transfected COS-7 and PAE cells, but not those which were not transfected, thereby establishing that ALK-1 is a receptor for TGF-ß.

This was confirmed in experiments on human umbilical vein endothelial cells (HUVEC). These cells are known to express ALK-1 endogenously, as well as ALK-5. The ALK-5 antiserum and the ALK-1 antiserum both immunoprecipitated type I and type II receptor cross linked complexes. The ALK-1 antiserum immunoprecipitated band migrated slightly more slowly than the band immunoprecipitated by the ALK-5 antiserum. This is in agreement with the difference in size of ALK-1 and ALK-5, and it indicates that both ALK-1 and ALK-5 bind TGF-S1 in HUVECS.

Further, it shows that ALK-1 acts as a co-called "type I" TGFß receptor in an endogenous, physiological setting.

Once it was determined that TGF-ß1 and ALK-1 interact, studies were carried out to determine whether or not activation of ALK-1 resulted in phosphorylation of Smads. To test this, COS-7 cells were transfected in the same manner described supra with either Flag tagged Smadl or Flag tagged Smad2 together with either a constitutively active form of ALK-1, or a constitutively active form of ALK-5. Specifically, the variant of ALK-1 is Q201D, and that of ALK-5 is T204D. Constitutively active ALK-1 was used to avoid the need for an additional transfection step. To elaborate, it is known that for the TGF-S pathway to function adequately, a complex of two, type I receptors, and two, type II receptors must interact, so as to activate the receptors. Constitutively active receptors, such as what was used herein, do not require the presence of the type II receptor to function. See Wieser et al., EMBO J 14: 2199-2208 (1995). In order to determine if the

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resulting transfected cells produced phosphorylated Smads, Smads were determined using a Flag specific antibody, which precipitated them, and phosphorylation was determined using antiphosphoserine antibody of Nishimura et al., J. Biol. Chem. 273: 1872-1879 (1998). It was determined, when the data were analyzed, that Smad1 was phosphorylated following interaction with activated ALK-1, but not following interaction of TGF-S the interaction of Conversely, TGF-S and ALK-5 led phosphorylation of Smad 2, but not Smad 1. This supports a conclusion that ALK-1 transduces signal in a manner similar to BMPs.

Additional experiments were then carried out to study the interaction of ALK-1 with Smad-1. Specifically, COS-7 cells were transfected with cDNA which encoded the wild type form of the TGF\$ type II receptor (TBR-II), a kinase inactive form of ALK-1, and Flag tagged Smad-1. Kinase inactive ALK-1 was used, because the interaction of Smad-1 and receptors is known to be transient, as once Smads are phosphorylated they dissociate from the type I receptor. See Marcias-Silva et al., Cell 87: 1215-1224 (1996); Nakao et al., EMBO J 16: 5353-5362 (1997). Affinity cross-linking, using 125I-TGF-\$1, and immunoprecipitation with Flag antibody was carried out, as discussed supra. The expression of ALK-1 was determined using anti-HA antibody, since the vector used to express ALK-1 effectively tagged it with HA.

The immunoprecipitating of Smadl resulted in coprecipitation of a cross linked TBR-II/ALK-1 complex, suggesting a direct association of Smadl with ALK-1.

These examples show that one can identify molecules which inhibit, or enhance expression of a gene whose expression is regulated by phosphorylated Smad1. To elaborate, as ALK-1 has been identified as a key constituent of the pathway by which Smad1 is phosphorylated, one can contact cells which express both Smad1 and ALK-1 with a substance of interest, and then determine if the Smad1 becomes phosphorylated. The cells can be those which inherently

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express both ALK-1 and Smadl, or which have been transformed or transfected with DNA encoding one or both of these. One can determine the phosphorylation via, e.g., the use of antiphosphorylated serine antibodies, as discussed supra. In an especially preferred embodiment, the assay can be carried out using TGF-ß, as a competing agent. The TGF-ß, as has been shown, does bind to ALK-1, leading to phosphorylation of Smadl. Hence, by determining a value with TGF-ß alone, one can then compare a value determined with amounts of the substance to be tested, in the presence of TGF-ß. Changes in phosphorylation levels can thus be attributed to the test substance.

In this type of system, it must be kept in mind that both type I receptors and type II receptors must be present; however, as indicated, supra, one can eliminate the requirement for a type II receptor by utilizing a constitutively active form of ALK-1, such as the form described supra. Additional approaches to inhibiting this system will be clear to the skilled artisan. For example, since it is known that there is interaction between Smad1 and the ALK-1 receptor, one can test for inhibition via the use of small molecules which inhibit the receptor/Smad interaction. al., supra, mention Smad6 and Smad7 as Smad1 inhibitors, albeit in the context of a different system. Hence one can test for inhibition, or inhibit the interaction, via adding a molecule to be tested or for actual inhibition to a cell, wherein the molecule is internalized by the cell, followed by assaying for phosphorylation, via a method such as is discussed supra.

In a similar way, one can assay for inhibitors of type I/type II receptor interaction, by testing the molecule of interest in a system which includes both receptors, and then assaying for phorphorylation.

Conversely, activators or agonists can also be tested for, or utilized, following the same type of procedures.

Via using any of these systems, one can identify any gene or genes which are activated by phosphorylated Smadl. To elaborate,

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the art is very familiar with systems of expression analysis, such as differential display PCR, subtraction hybridization, and other systems which combine driver and testes populations of nucleic acids, whereby transcripts which are expressed or not expressed can be identified. By simply using an activator/inhibitor of the system disclosed herein, on a first sample, and a second sample where none is used, one can then carry out analysis of transcript, thereby determining the transcripts of interest.

Also a part of the invention is the regulation of phosphorylation of Smad-1, with inhibitors, such as antibodies against the extracellular domain of ALK-1 or TGF-S, or enhancers, such as TGF-S itself, or those portions of the TGF-S molecule which are necessary for binding. Indeed, by appropriate truncation, one can also determine what portions of ALK-1 are required for phosphorylation of Smad1 to take place.

The invention has been described by way of example only, without restriction of its scope. The invention is defined by the subject matter herein, including the claims that follow the immediately following full Sequence Listings.

(1) GENERAL INFORMATION:

- (i) APPLICANTS: Miyazono, Kohei; Takeshe Imamura; ten Dijke, Peter
- (ii) TITLE OF INVENTION: Isolated ALK-1 Protein, Nucleic Acids Encoding It, And Uses Thereof
- (iii) NUMBER OF SEQUENCES: 29
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Felfe & Lynch
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 - (C) CITY: New York\City
 - (D) STATE: New York
 - (F) ZIP: 10022
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Diskette, 3.5 inch, 1.44 MB storage
 - (B) COMPUTER: IBM
 - (C) OPERATING SYSTEM: \ PC-DOS
 - (D) SOFTWARE: Wordper/fect
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: \08/436,265
 - (B) FILING DATE: 30-Octdber-1995
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: PCT/GB93/02367
 - (B) FILING DATE: 17-November-1993
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: 92/24057.1
 - (B) FILING DATE: 17-November-1992
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: 9304677.9
 - (B) FILING DATE: 8-March-1993
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: 9304680.3
 - (B) FILING DATE: 8-March-1993
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: 9311047.6
 - (B) FILING DATE: 28-May-1993

(vii) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: 9313763.6
(B) FILING DATE: 2-July-1993

<pre>(vii) PRIOR APPLICATION DATA: (A) APPLICATION NUMBER: 9136099.2 (B) FILING DATE: 3-August-1993</pre>	
<pre>(vii) PRIOR APPLICATION DATA: (A) APPLICATION NUMBER: 9321344.5 (B) FILING DATE: 15-October-1993</pre>	
<pre>(viii) ATTORNEY/AGENT INFORMATION: (A) NAME: Hanson, Norman D. (B) REGISTRATION NUMBER: 37,003 (C) REFERENCE/DOCKET NUMBER: LUD 5539</pre>	
(ix) TELECOMMUNICATION INFORMATION: (A) TELEPHONE: (212) 688-9200 (B) TELEFAX: (212) 838-3884	
(2) INFORMATION FOR SEQ ID NO: 1: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1984 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: unknown (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: NO (v) FRAGMENT TYPE: internal (vi) ORIGINAL SOURCE: (A) ORGANISM: Homo sapiens (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 2831791 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:	
AGGAAACGGT TTATTAGGAG GGAGTGGTGG AGCTGGGCCA GGCAGGAAGA CGCTGGAATA	60
AGAAACATTT TTGCTCCAGC CCCCATCCCA GTCCCGGGAG GCTGCCGCGC CAGCTGCGCC	120
GAGCGAGCCC CTCCCCGGCT CCAGCCCGGT CCGGGGCCGC GCCGGACCCC AGCCCGCCGT	180
CCAGCGCTGG CGGTGCAACT GCGGCCGCGC GGTGGAGGGG AGGTGGCCCC GGTCCGCCGA	240
AGGCTAGCGC CCCGCCACCC GCAGAGCGGG CCCAGAGGGA CC ATG ACC TTG GGC Met Thr Leu Gly 1	294

	CCC Pro																342
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	AGC Ser																438
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Cys	GGG Gly 70																534
©GTC	AAC Asn	CAC His	TAC Tyr	TGC Cys	TGC Cys 90	GAC Asp	AGC Ser	CAC His	CTC Leu	TGC Cys 95	AAC Asn	CAC His	AAC Asn	GTG Val	TCC Ser 100		582
CTG	GTG Val	CTG Leu	GAG Glu	GCC Ala 105	ACC Thr	CAA Gln	CCT Pro	CCT Pro	TCG Ser 110	GAG Glu	CAG Gln	CCG Pro	GGA Gly	ACA Thr 115	GAT Asp		630
GGC Gly	CAG Gln																678
GTG Val	GCC Ala	CTG Leu 135	GGT Gly	GTC Val	CTG Leu	GGC Gly	CTG Leu 140	TGG Trp	CAT His	GTC Val	CGA Arg	CGG Arg 145	AGG Arg	CAG Gln	GAG Glu		726
	CAG Gln 150																774
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							GAG Glu										1	L446
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	Thr	Cys	Thr 35	Cys	Glu	Ser	Pro	His 40	Cys	Lys	Gly	Pro	Thr 45	Cys	Arg	Gly	
	Ala	Trp 50	Cys	Thr	Val	Val	Leu 55	Val	Arg	Glu	Glu	Gly 60	Arg	His	Pro	Gln	

Glu His Arg Gly Cys Gly Asn Leu His Arg Glu Leu Cys Arg Gly Arg Pro Thr Glu Phe Val Asn His Tyr Cys Cys Asp Ser His Leu Cys Asn His Asn Val Ser Leu Val Leu Glu Ala Thr Gln Pro Pro Ser Glu Gln Pro Gly Thr Asp Gly Gln Leu Ala Leu Ile Leu Gly Pro Val Leu Ala Leu Leu Ala Leu Val Ala Leu Gly Val Leu Gly Leu Trp His Val Arg Arg Arg Gln Glu Lys Gln Arg Gly Leu His Ser Glu Leu Gly Glu Ser Ser Leu Ile Leu Lys Ala Ser Glu Gln Gly Asp Thr Met Leu Gly Asp Leu Leu Asp Ser Asp Cys Thr Thr Gly Ser Gly Ser Gly Leu Pro Phe Leu Val Gln Arg Thr Val Ala Arg Gln Val Ala Leu Val Glu Cys Val Gly Lys Gly Arg Tyr Gly Glu Val Trp Arg Gly Leu Trp His Gly Glu Ш Ser Val Ala Val Lys Ile Phe Ser Ser Arg Asp Glu Gln Ser Trp Phe Arg Glu Thr Glu Ile Tyr Asn Thr Val Leu Leu Arg His Asp Asn Ile Leu Gly Phe Ile Ala Ser Asp Met Thr Ser Arg Asn Ser Ser Thr Gln Leu Trp Leu Ile Thr His Tyr His Glu His Gly Ser Leu Tyr Asp Phe Leu Gln Arg Gln Thr Leu Glu Pro His Leu Ala Leu Arg Leu Ala Val Ser Ala Ala Cys Gly Leu Ala His Leu His Val Glu Ile Phe Gly Thr Gln Gly Lys Pro Ala Ile Ala His Arg Asp Phe Lys Ser Arg Asn Val

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Leu Val Lys Ser Asn Leu Gln Cys Cys Ile Ala Asp Leu Gly Leu Ala 340 345 350 Val Met His Ser Gln Gly Ser Asp Tyr Leu Asp Ile Gly Asn Asn Pro 355 Arg Val Gly Thr Lys Arg Tyr Met Ala Pro Glu Val Leu Asp Glu Gln Ile Arg Thr Asp Cys Phe Glu Ser Tyr Lys Trp Thr Asp Ile Trp Ala Phe Gly Leu Val Leu Trp Glu Ile Ala Arg Arg Thr Ile Val Asn Gly 405 415 Ile Val Glu Asp Tyr Arg Pro Pro Phe Tyr Asp Val Val Pro Asn Asp 425 Pro Ser Phe Glu Asp Met Lys Lys Val Val Cys Val Asp Gln Gln Thr Pro Thr Ile Pro Asn Arg Leu Ala Ala Asp Pro Val Leu Ser Gly Leu Ala Gln Met Met Arg Glu Cys Trp Tyr Pro Asn Pro Ser Ala Arg Leu **465** 470 475 480 Thr Ala Leu Arg Ile Lys Lys Thr Leu Gln Lys Ile Ser Asn Ser Pro 485 490 📒 Glu Lys Pro Lys Val Ile Gln 500

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	TGT Cys	GAA Glu	GGT Gly	CTC Leu 40	TCC Ser	TGC Cys	GGT Gly	AAT Asn	GAG Glu 45	GAC Asp	CAC His	TGT Cys	GAA Glu	GGC Gly 50	CAG Gln	CAG Gln	:	259
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					GTT Val													355
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	ACA Thr	CAG Gln	AAT Asn	TTC Phe 120	CAC His	TTG Leu	GAG Glu	GTT Val	GGC Gly 125	CTC Leu	ATT Ile	ATT Ile	CTC Leu	TCT Ser 130	GTA Val	GTG Val	4	99
	TTC Phe	GCA Ala	GTA Val 135	TGT Cys	CTT Leu	TTA Leu	GCC Ala	TGC Cys 140	CTG Leu	CTG Leu	GGA Gly	GTT Val	GCT Ala 145	CTC Leu	CGA Arg	AAA Lys	5	47
	TTT Phe	AAA Lys 150	AGG Arg	CGC Arg	AAC Asn	CAA Gln	GAA Glu 155	CGC Arg	CTC Leu	AAT Asn	CCC Pro	CGA Arg 160	GAC Asp	GTG Val	GAG Glu	TAT Tyr	5	95
	GGC Gly 165	ACT Thr	ATC Ile	GAA Glu	GGG Gly	CTC Leu 170	ATC Ile	ACC Thr	ACC Thr	AAT Asn	GTT Val 175	GGA Gly	GAC Asp	AGC Ser	ACT Thr	TTA Leu 180	6	343
	Ala	GAT Asp	TTA Leu	TTG Leu	GAT Asp 185	CAT His	TCG Ser	TGT Cys	ACA Thr	TCA Ser 190	GGA Gly	AGT Ser	GGC Gly	TCT Ser	GGT Gly 195	CTT Leu	6	591
	Pro	TTT Phe	CTG Leu	GTA Val 200	CAA Gln	AGA Arg	ACA Thr	GTG Val	GCT Ala 205	CGC Arg	CAG Gln	ATT Ile	ACA Thr	CTG Leu 210	TTG Leu	GAG Glu	7	739
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· C	Gly	GAA Glu 230	AAT Asn	GTT Val	GCC Ala	GTG Val	AAG Lys 235	ATC Ile	TTC Phe	TCC Ser	TCC Ser	CGT Arg 240	GAT Asp	GAG Glu	AAG Lys	TCA Ser	8	335
1	TGG	Phe	AGG Arg	GAA Glu	ACG Thr	GAA Glu 250	TTG Leu	TAC Tyr	AAC Asn	ACT Thr	GTG Val 255	ATG Met	CTG Leu	AGG Arg	CAT His	GAA Glu 260	8	883
	AAT Asn	ATC Ile	TTA Leu	GGT Gly	TTC Phe 265	ATT Ile	GCT Ala	TCA Ser	GAC Asp	ATG Met 270	ACA Thr	TCA Ser	AGA Arg	CAC His	TCC Ser 275	AGT Ser	9	931
	ACC Thr	CAG Gln	CTG Leu	TGG Trp 280	TTA Leu	ATT Ile	ACA Thr	CAT His	TAT Tyr 285	CAT His	GAA Glu	ATG Met	GGA Gly	TCG Ser 290	Leu	TAC Tyr	•	979
	GAC Asp	TAT Tyr	CTT Leu 295	CAG Gln	CTT Leu	ACT Thr	ACT Thr	CTG Leu 300	GAT Asp	ACA Thr	GTT Val	AGC Ser	TGC Cys 305	CTT Leu	CGA Arg	ATA Ile	10	027
•	GTG Val	CTG Leu 310	Ser	ATA Ile	GCT Ala	AGT Ser	GGT Gly 315	Leu	GCA Ala	CAT His	TTG Leu	CAC His 320	Ile	GAG Glu	ATA Ile	TTT Phe	1	075

		ACC Thr															1123
		ATT Ile															1171
		GCA Ala															1219
		CCC Pro															1267
	_	ACC Thr 390															1315
41	TGG Trp 405	GCC Ala	TTT Phe	GGA Gly	CTT Leu	GTT Val 410	TTG Leu	TGG Trp	GAA Glu	GTG Val	GCC Ala 415	AGG Arg	CGG Arg	ATG Met	GTG Val	AGC Ser 420	1363
<u>i</u>		GGT Gly															1411
	AAT Asn	GAC Asp	CCA Pro	AGT Ser 440	TTT Phe	GAA Glu	GAT Asp	ATG Met	AGG Arg 445	AAG Lys	GTA Val	GTC Val	TGT Cys	GTG Val 450	GAT Asp	CAA Gln	1459
	CAA Gln	AGG Arg	CCA Pro 455	AAC Asn	ATA Ile	CCC Pro	AAC Asn	AGA Arg 460	TGG Trp	TTC Phe	TCA Ser	GAC Asp	CCG Pro 465	ACA Thr	TTA Leu	ACC Thr	1507
		CTG Leu 470															1555
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	ACTGGGCATT	TCACGAACTG	TTCACACTGC	AGAGACTAAT	GTTGGACAGA	CACTGTTGCA	1890
	AAGGTAGGGA	CTGGAGGAAC	ACAGAGAAAT	CCTAAAAGAG	ATCTGGGCAT	TAAGTCAGTG	1950
	GCTTTGCATA	GCTTTCACAA	GTCTCCTAGA	CACTCCCCAC	GGGAAACTCA	AGGAGGTGGT	2010
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	ATGTTTTAA	CACTATACTC	TAAAATGGAC	ATTTTCTTTT	ATTATCAGTT	AAAATCACAT	2430
	TTTAAGTGCT	TCACATTTGT	ATGTGTGTAG	ACTGTAACTT	TTTTTCAGTT	CATATGCAGA	2490
ļ	ACGTATTTAG	CCATTACCCA	CGTGACACCA	CCGAATATAT	TATCGATTTA	GAAGCAAAGA	2550
.044.	•	ATTTTAGTCC	TGAACGCTAC	GGGGAAAATG	CATTTTCTTC	AGAATTATCC	2610
a 1111	ATTACGTGCA	TTTAAACTCT	GCCAGAAAA	AATAACTATT	TTGTTTTAAT	CTACTTTTTG	2670
		TTATTTGTAT	AAATTAAATA	AACTGTTTTC	AAGTCAAAAA	AAAA	2724
F. F.							
ū		MATION FOR S	SEQ ID NO: 4				the fire and Assessing spiritures and

(2) INFORMATION FOR SEQ ID NO: 4:

- SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 509 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

Met Val Asp Gly Val Met Ile Leu Pro Val Leu Ile Met Ile Ala Leu 10

Pro Ser Pro Ser Met Glu Asp Glu Lys Pro Lys Val Asn Pro Lys Leu 20 25 30

Tyr Met Cys Val Cys Glu Gly Leu Ser Cys Gly Asn Glu Asp His Cys 35 40 45

Glu Gly Gln Gln Cys Phe Ser Ser Leu Ser Ile Asn Asp Gly Phe His Val Tyr Gln Lys Gly Cys Phe Gln Val Tyr Glu Gln Gly Lys Met Thr 65 Cys Lys Thr Pro Pro Ser Pro Gly Gln Ala Val Glu Cys Cys Gln Gly Asp Trp Cys Asn Arg Asn Ile Thr Ala Gln Leu Pro Thr Lys Gly Lys Ser Phe Pro Gly Thr Gln Asn Phe His Leu Glu Val Gly Leu Ile Ile 115 120 125 Leu Ser Val Val Phe Ala Val Cys Leu Leu Ala Cys Leu Leu Gly Val Ala Leu Arg Lys Phe Lys Arg Arg Asn Gln Glu Arg Leu Asn Pro Arg Asp Val Glu Tyr Gly Thr Ile Glu Gly Leu Ile Thr Thr Asn Val Gly 170 Asp Ser Thr Leu Ala Asp Leu Leu Asp His Ser Cys Thr Ser Gly Ser 180 Gly Ser Gly Leu Pro Phe Leu Val Gln Arg Thr Val Ala Arg Gln Ile 195 200 High Thr Leu Leu Glu Cys Val Gly Lys Gly Arg Tyr Gly Glu Val Trp Arg 🏢 Gly Ser Trp Gln Gly Glu Asn Val Ala Val Lys Ile Phe Ser Ser Arg 225 235 240 Asp Glu Lys Ser Trp Phe Arg Glu Thr Glu Leu Tyr Asn Thr Val Met 245 250 255 Leu Arg His Glu Asn Ile Leu Gly Phe Ile Ala Ser Asp Met Thr Ser Arg His Ser Ser Thr Gln Leu Trp Leu Ile Thr His Tyr His Glu Met 280 -285 Gly Ser Leu Tyr Asp Tyr Leu Gln Leu Thr Thr Leu Asp Thr Val Ser 290 295 300 Cys Leu Arg Ile Val Leu Ser Ile Ala Ser Gly Leu Ala His Leu His 305 310 315 320

Ile Glu Ile Phe Gly Thr Gln Gly Lys Pro Ala Ile Ala His Arg Asp 325 330 Leu Lys Ser Lys Asn Ile Leu Val Lys Lys Asn Gly Gln Cys Cys Ile 345 Ala Asp Leu Gly Leu Ala Val Met His Ser Gln Ser Thr Asn Gln Leu 360 Asp Val Gly Asn Asn Pro Arg Val Gly Thr Lys Arg Tyr Met Ala Pro Glu Val Leu Asp Glu Thr Ile Gln Val Asp Cys Phe Asp Ser Tyr Lys 385 390 Arg Val Asp Ile Trp Ala Phe Gly Leu Val Leu Trp Glu Val Ala Arg 405 410 Arg Met Val Ser Asn Gly Ile Val Glu Asp Tyr Lys Pro Pro Phe Tyr Asp Val Val Pro Asn Asp Pro Ser Phe Glu Asp Met Arg Lys Val Val 435 440 🗎 Cys Val Asp Gln Gln Arg Pro Asn Ile Pro Asn Arg Trp Phe Ser Asp 450 455 Pro Thr Leu Thr Ser Leu Ala Lys Leu Met Lys Glu Cys Trp Tyr Gln **465** 470 W 🚉 Asn Pro Ser Ala Arg Leu Thr Ala Leu Arg Ile Lys Lys Thr Leu Thr 490 E Lys Ile Asp Asn Ser Leu Asp Lys Leu Lys Thr Asp Cys

500

(2)	(i. (i. (v. (v. (ix.) Si (i (i) (i ii) Mo iii) I v) Al (i) FEI (i) (i)	ATION EQUEN A) LI B) T C) S C) S C) T C) LC C C C C C C C C C C C C C C C C C C	NCE (ENGT) YPE: FRANI OPOLO ULE ! FENSI NT T NAL ! RGAN: E: AME/I OCAT:	CHARA H: 29 nucl DEDNI DGY: IYPE: ICAL E: NO YPE: SOURC ISM: KEY: ION:	ACTED 32 1 leic ESS: line CDD inte CCE: Homo	RIST: pase acid unkrear NA erna: p sag	ICS: pair d nown	3		-					
	(x:	i) S	EQUE	NCE 1	DESCI	RIPT:	ION:	SEQ	ID 1	10: !	5:					
GC:	rccgc	GCC (GAGG	GCTG(GA G	GATG(CGTT	c cc:	rggg	GTCC	GGA	CTTAT	rga i	AAAT	ATGCAT	60
CAC	STTTA	ATA (CTGT	CTTG	GA A	rtca:	rgagi	A TG	GAAG	CATA	GGT	CAAA	GCT (GTTT(GAGAA	120
	CAGA	AGT I	ACAG:	r t tt	AT C	ragco	CACA:	r ct	rgga	GGAG	TCG	raag <i>i</i>	AAA (GCAGT	rgggag	180
	GAAGT	CAT :	TGTC2	AAGT	GC T	rgcgz	ATCT:	r tt	ACAA	GAAA	ATC	rcac:	rga i	ATGAT	TAGTCA	240
TT:	TAAAT	TGG :	rgaa(GTAG(CA A	GACC!	AATT	A TT	AAAG	GTGA	CAG	raca(CAG (GAAA(CATTAC	300
AA!	rtgaa(IG A(et T) 1								rg Le					348
U TAT	TTG Leu 15	Phe													ATG Met	396
	CAT His															444
	GGA Gly															492
	TGC Cys															540
	T AAT Asn															588

.

							TGT Cys 100										636
C							GCC Ala										684
							CAG Gln										732
_							GAT Asp										780
	le	Ser	Met 160	Ala	Val	Cys		Ile 165	Ala	Met	Ile	Ile	Phe 170	Ser	Ser	Сув	828
<u></u> 1	TTT Phe	TGT Cys 175	TAC Tyr	AAA Lys	CAT His	TAT Tyr	TGC Cys 180	AAG Lys	AGC Ser	ATC Ile	TCA Ser	AGC Ser 185	AGA Arg	CGT Arg	CGT Arg	TAC Tyr	876
7	AAT Asn L90	CGT Arg	GAT Asp	TTG Leu	GAA Glu	CAG Gln 195	GAT Asp	GAA Glu	GCA Ala	TTT Phe	ATT Ile 200	CCA Pro	GTT Val	GGA Gly	GAA Glu	TCA Ser 205	924
							CAG Gln										972
							CGA Arg										1020
							CGA Arg										1068
							GTG Val 260										1116
S							GAA Glu										1164
							ATA Ile										1212

	ACT															1260
															AAA Lys	1308
	GCT Ala 335															1356
	GGC Gly															 1404
Lys	AAC Asn	Ile	Leu	Ile 370	Lys	Lys	Asn	Gly	Ser 375	Cys	Cys	Ile	Ala	Asp 380	Leu	1452
OGGC Gly	CTT Leu	GCT Ala	GTT Val 385	AAA Lys	TTC Phe	AAC Asn	AGT Ser	GAC Asp 390	ACA Thr	AAT Asn	GAA Glu	GTT Val	GAT Asp 395	GTG Val	CCC Pro	1500
TTG	AAT Asn	ACC Thr 400	AGG Arg	GTG Val	GGC Gly	ACC Thr	AAA Lys 405	CGC Arg	TAC Tyr	ATG Met	GCT Ala	CCC Pro 410	GAA Glu	GTG Val	CTG Leu	1548
GAC W Asp																1596
TATC TILE 430																1644
	GGA Gly															1692
	AGT Ser															1740
	TTG Leu															1788
	GCA Ala 495															1836

	TCC F Ser F 510																	1884
	GAA T							TGAT	'GGT'	TAA Z	acca1	rcgg <i>i</i>	AG G	AGAA	ACTCI	ŗ		1935
	AGACI	rgca	AG.	AACT	STTTI	T A	CCAI	rggca	TGC	GTG	GAAT	TAGA	GTG	SAA	TAAGO	SATGT	T	1995
	AACTI	rggī	TC '	TCAG	ACTCI	T T	CTTC	ACTAC	GTO	TTC	ACAG	GCT	CTA	ATA	TTAAI	ACCTT	T	2055
	CAGTA	ACTO	TT .	ATTA	GGAT <i>A</i>	C A	AGCTO	GGAA	CTI	CTA	AACA	CTTC	CATTO	CTT	TATAT	ratgg	A	2115
	CAGCI	ATT	TT	TTAAI	ATGT	G T	TTTT	ATGC	CT	TTTT:	TAA	GTG	GTTI	TT	ATGA	ACTGC	A	2175
	TCAAG	BACT	TC .	AATC	CTGAI	T A	STGTO	CTCCA	GTO	CAAG	CTCT	GGGI	ACT	BAA	TTGC	CTGTT	С	2235
=	ATAAA	AACG	GT (GCTT	rctgi	G A	AAGCC	CTTAA	GAZ	\GAT	TAAA	GAGO	GCAG	GCA	GAGAI	rggag	A	2295
	AATAG	ACT	TT (GCCTI	CATTI	C TO	GAGAC	CATTO	AG1	TCG:	rttg	TATI	CTAC	CCT	TTGT	AAAAC	A	2355
	GCCTA	ATAG	AT (GATG!	ATGT	T T	rggg <i>i</i>	ATACI	GCI	TAT?	TTTA	TGAI	AGTI	TG	TCCT	STGTC	C	2415
See. Or	TTAGI	GAT	GT (GTGT	STGTC	T C	CATGO	CACAT	GCI	ACGC	CGGG	ATTO	CTCI	rgc	TGCC	ATTTG	A	2475
4	ATTAG	BAAG	AA .	AATAA	ATTTA	T AT	rgcai	rgcac	AGO	BAAG	ATAT	TGGI	GGCC	CGG	TGGTI	TTGT	G.	2535
the Healt	CTTTA	AAA	AT (GCAA1	TATCI	G A	CCAAG	ATTO	GCC	CAAT	CTCA	TACA	AGCC	CAT	TTACI	TTGC	A	2595
	AGTGA	GAT	'AG	CTTC	CCCAC	C A	GCTT1	TTTAT	TTI	TAAC	ATGA	AAGO	TGAT	rgc	CAAGO	CCAA	A	2655
4	AGAAG	TTT	'AA	AGCAT	rctgi	'A A	ATTTO	GACT	GTI	TTC	CTTC	AACC	CACCA	TT	TTTTI	TGTG	G	2715
	TTATI	TTA	TT '	rgtc <i>i</i>	ACGGA	A A	CATO	CCTCT	CCZ	AAG:	rtgg	AGCI	TCT	ATT	GCCAI	rgaac	С	2775
	ATGCT	TAC	AA .	AGAAZ	AGCAC	T TO	CTTAT	TGAA	GTO	AAT	rcct	GCAT	TTG	ATA	GCAA]	rgtaa	G	2835
	TGCCI	ATA	AC (CATGI	TCTA	T A	TCTI	TATT	CTC	CAGT	AACT	TTTA	ÄAAG	GG	AAGTI	TTTAT	A	2895
	TATTI	TGT	GT 2	CAATA	rgrgc	T T	rttan	GCAA	ATO	CACC	C							2932

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- (2) INFORMATION FOR SEQ ID NO: 6:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 532 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

Met Thr Gln Leu Tyr Ile Tyr Ile Arg Leu Leu Gly Ala Tyr Leu Phe 1 5 10 15

Ile Ile Ser Arg Val Gln Gly Gln Asn Leu Asp Ser Met Leu His Gly
20 25 30

Thr Gly Met Lys Ser Asp Ser Asp Gln Lys Lys Ser Glu Asn Gly Val 35 40 45

Thr Leu Ala Pro Glu Asp Thr Leu Pro Phe Leu Lys Cys Tyr Cys Ser 50 60

Gly His Cys Pro Asp Asp Ala Ile Asn Asn Thr Cys Ile Thr Asn Gly 65 70 75 80

His Cys Phe Ala Ile Ile Glu Glu Asp Asp Gln Gly Glu Thr Thr Leu 85 90 95

Ala Ser Gly Cys Met Lys Tyr Glu Gly Ser Asp Phe Gln Cys Lys Asp 100 105 110

Ser Pro Lys Ala Gln Leu Arg Arg Thr Ile Glu Cys Cys Arg Thr Asn 115 120 125

Leu Cys Asn Gln Tyr Leu Gln Pro Thr Leu Pro Pro Val Val Ile Gly

Pro Phe Phe Asp Gly Ser Ile Arg Trp Leu Val Leu Leu Ile Ser Met 145 150 155 160

Ala Val Cys Ile Ile Ala Met Ile Ile Phe Ser Ser Cys Phe Cys Tyr 165 170 175

Lys His Tyr Cys Lys Ser Ile Ser Ser Arg Arg Arg Tyr Asn Arg Asp 180 185 190

Leu Glu Gln Asp Glu Ala Phe Ile Pro Val Gly Glu Ser Leu Lys Asp 195 200 205

Leu Ile Asp Gln Ser Gln Ser Ser Gly Ser Gly Leu Pro Leu 210 215 220

Leu Val Gln Arg Thr Ile Ala Lys Gln Ile Gln Met Val Arg Gln Val Gly Lys Gly Arg Tyr Gly Glu Val Trp Met Gly Lys Trp Arg Gly Glu Lys Val Ala Val Lys Val Phe Phe Thr Thr Glu Glu Ala Ser Trp Phe Arg Glu Thr Glu Ile Tyr Gln Thr Val Leu Met Arg His Glu Asn Ile Leu Gly Phe Ile Ala Ala Asp Ile Lys Gly Thr Gly Ser Trp Thr Gln Leu Tyr Leu Ile Thr Asp Tyr His Glu Asn Gly Ser Leu Tyr Asp Phe Leu Lys Cys Ala Thr Leu Asp Thr Arg Ala Leu Leu Lys Leu Ala Tyr Ser Ala Ala Cys Gly Leu Cys His Leu His Thr Glu Ile Tyr Gly Thr Gln Gly Lys Pro Ala Ile Ala His Arg Asp Leu Lys Ser Lys Asn Ile Leu Ile Lys Lys Asn Gly Ser Cys Cys Ile Ala Asp Leu Gly Leu Ala Hal Lys Phe Asn Ser Asp Thr Asn Glu Val Asp Val Pro Leu Asn Thr 🏥 Arg Val Gly Thr Lys Arg Tyr Met Ala Pro Glu Val Leu Asp Glu Ser Leu Asn Lys Asn His Phe Gln Pro Tyr Ile Met Ala Asp Ile Tyr Ser Phe Gly Leu Ile Ile Trp Glu Met Ala Arg Arg Cys Ile Thr Gly Gly Ile Val Glu Glu Tyr Gln Leu Pro Tyr Tyr Asn Met Val Pro Ser Asp Pro Ser Tyr Glu Asp Met Arg Glu Val Val Cys Val Lys Arg Leu Arg Pro Ile Val Ser Asn Arg Trp Asn Ser Asp Glu Cys Leu Arg Ala Val

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Leu Lys Leu Met Ser Glu Cys Trp Ala His Asn Pro Ala Ser Arg Leu

Thr Ala Leu Arg Ile Lys Lys Thr Leu Ala Lys Met Val Glu Ser Gln

Asp Val Lys Ile

			INFORMATION FOR SEQ ID NO: 7: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2333 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: unknown (D) TOPOLOGY: linear (ii) MOLECULE TYPE: CDNA (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: NO (v) FRAGMENT TYPE: internal (vi) ORIGINAL SOURCE: (A) ORGANISM: Homo sapiens (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 11515 xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7: CG GAG TCG GCC GGA GCC TCC TCC TTC TTC CCC CTT GTT GTC CTC la Glu Ser Ala Gly Ala Ser Ser Phe Phe Pro Leu Val Val Leu 5 IC GCC GGC AGC GGC GGG TCC GGG CCC CGG GGG GTC CAG GCT CTG eu Ala Gly Ser Gly Gly Ser Gly Pro Arg Gly Val Gln Ala Leu 20 25 GT GCG TGC ACC AGC TGC CTC CAG GCC AAC TAC ACG TGT GAG ACA 144 ys Ala Cys Thr Ser Cys Leu Gln Ala Asn Tyr Thr Cys Glu Thr 35 40 45														
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		(i:	•			vev.	ana										
A. III					-			1515									
		(xi)							SEQ :	ID NO	0: 7	:					٠
_;																	
	ATG	GCG	GAG	TCG	GCC	GGA	GCC	TCC	TCC	TTC	TTC	CCC	CTT	GTT	GTC	CTC	48
ٔ لِيا	1	ALG	GIU	SET	_	GLY	ALG	Ser	per		FITE	PIO	пеп	vaı		neu	
Ū																	
IJ	CTG	CTC	GCC	GGC	AGC	GGC	GGG	TCC	GGG	CCC	CGG	GGG	GTC	CAG	GCT	CTG	96
	Leu	гел	Ата		ser	GIA	GIY	ser		Pro	Arg	GIY	vaı		Ala	Leu	
				20					23					30			
																	144
	Leu	Cys		Cys	Thr	Ser	Cys		Gln	Ala	Asn	Tyr		Cys	Glu	Thr	
			33					7.0					43				
				TGC													192
	Asp		Ala	Cys	Met	Val		Phe	Phe	Asn	Leu		Gly	Met	Glu	His	
		50					55		•			60					
	CAT	GTG	CGC	ACC	TGC	ATC	CCC	AAA	GTG	GAG	CTG	GTC	CCT	GCC	GGG	AAG	240
1		Val	Arg	Thr	Cys		Pro	Lys	Val	Glu		Val	Pro	Ala	Gly	_	
	65					70					75					80	

		TTC Phe															288
		ACT Thr															336
		AAG Lys															384
		ATC Ile 130				Pro											432
	Val 145	TTC Phe															480
	AGA Arg	CTG Leu															528
	AAG	ACG Thr														GGC Gly	576
	TCA Ser	GGG Gly	TTA Leu 195	CCC Pro	CTC Leu	TTT Phe	GTC Val	CAG Gln 200	CGC Arg	ACA Thr	GTG Val	GCC Ala	CGA Arg 205	ACC Thr	ATC Ile	GTT Val	624
2 : :		CAA Gln 210															672
		TGG Trp															720
		CGG Arg															768
		CAT His															816
		ACC Thr															864

						AAC Asn 295										912
						GCT Ala										960
						GGG Gly										1008
						GTG Val										1056
						CGT Arg										1104
EATT Ile	GCC Ala 370	CCG Pro	AAT Asn	CAG Gln	AGG Arg	GTG Val 375	GGG Gly	ACC Thr	AAA Lys	CGA Arg	TAC Tyr 380	ATG Met	GCC Ala	CCT Pro	GAA Glu	1152
						AAT Asn										1200
☐ GCT ₩ Ala ⊭																1248
Ū TGC Ū Cys																1296
						TCC Ser										1344
						AAC Asn 455										1392
						AAG Lys										1440
						GCC Ala										1488

	G CAG GAA GA l Gln Glu Aa 500	sp Val Lys	ATC TAACTGC Ile 505	FCC CTCTCTCC	CAC	1535
ACGGAGCTCC	TGGCAGCGAG	AACTACGCAC	AGCTGCCGCG	TTGAGCGTAC	GATGGAGGCC	1595
TACCTCTCGT	TTCTGCCCAG	CCCTCTGTGG	CCAGGAGCCC	TGGCCCGCAA	GAGGGACAGA	1655
GCCCGGGAGA	GACTCGCTCA	CTCCCATGTT	GGGTTTGAGA	CAGACACCTT	TTCTATTTAC	1715
CTCCTAATGG	CATGGAGACT	CTGAGAGCGA	ATTGTGTGGA	GAACTCAGTG	CCACACCTCG	1775
AACTGGTTGT	AGTGGGAAGT	CCCGCGAAAC	CCGGTGCATC	TGGCACGTGG	CCAGGAGCCA	1835
TGACAGGGGC	GCTTGGGAGG	GGCCGGAGGA	ACCGAGGTGT	TGCCAGTGCT	AAGCTGCCCT	1895
GAGGGTTTCC	TTCGGGGACC	AGCCCACAGC	ACACCAAGGT	GGCCCGGAAG	AACCAGAAGT	1955
GCAGCCCCTC	TCACAGGCAG	CTCTGAGCCG	CGCTTTCCCC	TCCTCCCTGG	GATGGACGCT	2015
GCCGGGAGAC	TGCCAGTGGA	GACGGAATCT	GCCGCTTTGT	CTGTCCAGCC	GTGTGTGCAT	2075
GTGCCGAGGT	GCGTCCCCCG	TTGTGCCTGG	TTCGTGCCAT	GCCCTTACAC	GTGCGTGTGA	2135
GTGTGTGTGT	GTGTCTGTAG	GTGCGCACTT	ACCTGCTTGA	GCTTTCTGTG	CATGTGCAGG	2195
TCGGGGGTGT	GGTCGTCATG	CTGTCCGTGC	TTGCTGGTGC	CTCTTTTCAG	TAGTGAGCAG	2255
CATCTAGTTT	CCCTGGTGCC	CTTCCCTGGA	GGTCTCTCCC	TCCCCCAGAG	CCCCTCATGC	2315
CACAGTGGTA	CTCTGTGT					2333

- (2) INFORMATION FOR SEQ ID NO: 8:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 505 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

Met Ala Glu Ser Ala Gly Ala Ser Ser Phe Phe Pro Leu Val Val Leu
1 5 10 15

Leu Leu Ala Gly Ser Gly Gly Ser Gly Pro Arg Gly Val Gln Ala Leu 20 25 30

Leu Cys Ala Cys Thr Ser Cys Leu Gln Ala Asn Tyr Thr Cys Glu Thr 35 40 45

Asp Gly Ala Cys Met Val Ser Phe Phe Asn Leu Asp Gly Met Glu His His Val Arg Thr Cys Ile Pro Lys Val Glu Leu Val Pro Ala Gly Lys Pro Phe Tyr Cys Leu Ser Ser Glu Asp Leu Arg Asn Thr His Cys Cys Tyr Thr Asp Tyr Cys Asn Arg Ile Asp Leu Arg Val Pro Ser Gly His Leu Lys Glu Pro Glu His Pro Ser Met Trp Gly Pro Val Glu Leu Val 120 115 125 Gly Ile Ile Ala Gly Pro Val Phe Leu Phe Leu Ile Ile Ile Ile 135 140 Val Phe Leu Val Ile Asn Tyr His Gln Arg Val Tyr His Asn Arg Gln Arg Leu Asp Met Glu Asp Pro Ser Cys Glu Met Cys Leu Ser Lys Asp 165 170 Lys Thr Leu Gln Asp Leu Val Tyr Asp Leu Ser Thr Ser Gly Ser Gly 180 185 190 Ser Gly Leu Pro Leu Phe Val Gln Arg Thr Val Ala Arg Thr Ile Val U Heu Gln Glu Ile Ile Gly Lys Gly Arg Phe Gly Glu Val Trp Arg Gly 🏥 Arg Trp Arg Gly Gly Asp Val Ala Val Lys Ile Phe Ser Ser Arg Glu 225 230 235 240 Glu Arg Ser Trp Phe Arg Glu Ala Glu Ile Tyr Gln Thr Val Met Leu 245 250 255 Arg His Glu Asn Ile Leu Gly Phe Ile Ala Ala Asp Asn Lys Asp Asn Gly Thr Trp Thr Gln Leu Trp Leu Val Ser Asp Tyr His Glu His Gly 275 280 285 Ser Leu Phe Asp Tyr Leu Asn Arg Tyr Thr Val Thr Ile Glu Gly Met 290 295 300 Ile Lys Leu Ala Leu Ser Ala Ala Ser Gly Leu Ala His Leu His Met 305 310 315

Glu Ile Val Gly Thr Gln Gly Lys Pro Gly Ile Ala His Arg Asp Leu 325 Lys Ser Lys Asn Ile Leu Val Lys Lys Asn Gly Met Cys Ala Ile Ala 340 Asp Leu Gly Leu Ala Val Arg His Asp Ala Val Thr Asp Thr Ile Asp Ile Ala Pro Asn Gln Arg Val Gly Thr Lys Arg Tyr Met Ala Pro Glu 375 Val Leu Asp Glu Thr Ile Asn Met Lys His Phe Asp Ser Phe Lys Cys 390 385 400 Ala Asp Ile Tyr Ala Leu Gly Leu Val Tyr Trp Glu Ile Ala Arg Arg 405 Cys Asn Ser Gly Gly Val His Glu Glu Tyr Gln Leu Pro Tyr Tyr Asp 425 Leu Val Pro Ser Asp Pro Ser Ile Glu Glu Met Arg Lys Val Val Cys Asp Gln Lys Leu Arg Pro Asn Ile Pro Asn Trp Trp Gln Ser Tyr Glu 450 Ala Leu Arg Val Met Gly Lys Met Met Arg Glu Cys Trp Tyr Ala Asn 470 Gly Ala Ala Arg Leu Thr Ala Leu Arg Ile Lys Lys Thr Leu Ser Gln 485 490 🛱 Leu Ser Val Gln Glu Asp Val Lys Ile 500 505

(2)	(i) (ii (iv (v) (vi	(1 (1 (1) M(1 (1) H(1) H(1) H(1) H(1) (1) FR4 (1) (1) (1) (1) (1) (1) (1)	EQUEN () LE () SI () SI () TO () TO () TO () C () O () O () O () O () O () O () O () O	ICE OF THE TOTAL CONTROL OF THE TOTAL SENSE OF THE	CHARA I: 23 nucl DEDNE OGY: CYPE: COAL: COM: COM:	CTER 08 H eic SS: line CDN inte Mous CDS 77	RISTI pase acid unkrear NA ernal	CCS: pair l lown		10: S):						
	GAGG	CGA (GTTI	GCT	G GC	TGAC	GCA	G CGC	CGCC	GCC	GGG	CCGGG	CC C	GGC	CACAGO	3 6	0
CGC CGC CGC	TGGC	GGC (GGA(CC AT	rg g <i>i</i> et g] 1	AG GC Lu Al	CG GC la Al	CG GI la Va	C GC Al Al 5	CT GC la Al	CT CC la Pi	CG CG	g Pı	CC CC CO A1 LO	eg eg	10	19
E CTO	G CTC	CTC Leu	CTC Leu 15	GTG Val	CTG Leu	GCG Ala	GCG Ala	GCG Ala 20	GCG Ala	GCG Ala	GCG Ala	GCG Ala	GCG Ala 25	GCG Ala	CTG Leu	15	; 7
լլ Leւ ⊭	C CCG	Gly 30	Ala	Thr	Ala	Ļeu	Gln 35	Cys	Phe	Cys	His	Leu 40	Cys	Thr	Lys	20	15
GAC G Asi	C AAT Asn 45	TTT Phe	ACT Thr	TGT Cys	GTG Val	ACA Thr 50	nop	GGG Gly	200	TGC Cys	TTT Phe 55	GTC Val	TCT Ser	GTC Val	ACA Thr	25	3
GAC Glu	ACC Thr	ACA Thr	GAC Asp	AAA Lys	GTT Val 65	ATA Ile	CAC His	AAC Asn	AGC Ser	ATG Met 70	TGT Cys	ATA Ile	GCT Ala	GAA Glu	ATT Ile 75	30)1
GA(As _I	C TTA D Leu	ATT Ile	CCT Pro	CGA Arg 80	GAT Asp	AGG Arg	CCG Pro	TTT	GTA Val 85	TGT Cys	GCA Ala	CCC Pro	TCT Ser	TCA Ser 90	AAA Lys	34	19
AC.	r GGG r Gly	TCT Ser	GTG Val 95	ACT Thr	ACA Thr	ACA Thr	TAT Tyr	TGC Cys 100	TGC Cys	AAT Asn	CAG Gln	GAC Asp	CAT His 105	TGC	AAT Asn	39	₹7
AA Lys	A ATA 3 Ile	GAA Glu 110	CTT Leu	CCA Pro	ACT Thr	ACT Thr	GTA Val 115	AAG Lys	TCA Ser	TCA Ser	CCT Pro	GGC Gly 120	CTT Leu	GGT Gly	CCT Pro	44	15

	GAA L Glu 125															493
	A CTC c Leu															541
	CGA Arg															589
	A GAG															637
	TCT Ser															685
1 2	T ATT	Val														733
	AGA Arg															781
TC: USe: U	AGA Arg	GAA Glu	GAA Glu	CGT Arg 240	TCG Ser	TGG Trp	TTC Phe	CGT Arg	GAG Glu 245	GCA Ala	GAG Glu	ATT Ile	TAT Tyr	CAA Gln 250	ACT Thr	829
	A ATG L Met													Asp		877
	A GAC s Asp															925
	CAT His 285															973
	A GGA 1 Gly)															1021
	CAC His															1069

														GGA Gly 345			1117
														GCC Ala			1165
														AGG Arg			1213
														TTT Phe			1261
	Phe													TGG Trp	_		1309
	Ala	CGA Arg	CGA Arg	TGT Cys 415	TCC Ser	ATT Ile	GGT Gly	GGA Gly	ATT Ile 420	CAT His	GAA Glu	GAT Asp	TAC Tyr	CAA Gln 425	CTG Leu	CCT Pro	1357
~===														ATG Met		AAA Lys	1405
														AGA Arg			1453
														GAA Glu		TGG Trp 475	1501
														AAG Lys			1549
							CAG Gln						TAAT	TTCT?	ACA		1595
	GCTI	TGCC	CTG A	ACTO	CTCCI	T TI	TTCI	TCAC	ATC	TGCI	CCT	GGGI	TTT	L TA	TGGC	AGGTC	1655
	AGTI	GTTC	TA C	CTCA	CTG	AG AG	GGAA	CAGA	AGG	ATAI	TGC	TTCC	TTT	rgc <i>i</i>	GCA	TGTAA	1715
	TAAA	GTCA	AT I	AAAA	ACTI	יכ ככ	CAGGA	TTTC	TTI	GGAC	CCA	GGAA	ACAC	SCC A	TGT	GGTCC	1775
	TTTC	TGTG	CA C	CTATO	AACO	C TI	CTTI	CCCA	GGA	CAGA	AAA	TGT	TAGI	CT A	CCTI	TTATT	1835

TTTATTAACA	AAACTTGTTT	TTTAAAAAGA	TGATTGCTGG	TCTTAACTTT	AGGTAACTCT	1895
GCTGTGCTGG	AGATCATCTT	TAAGGGCAAA	GGAGTTGGAT	TGCTGAATTA	CAATGAAACA	1955
TGTCTTATTA	CTAAAGAAAG	TGATTTACTC	CTGGTTAGTA	CATTCTCAGA	GGATTCTGAA	2015
CCACTAGAGT	TTCCTTGATT	CAGACTTTGA	ATGTACTGTT	CTATAGTTTT	TCAGGATCTT	2075
AAAACTAACA	CTTATAAAAC	TCTTATCTTG	AGTCTAAAAA	TGACCTCATA	TAGTAGTGAG	2135
GAACATAATT	CATGCAATTG	TATTTTGTAT	ACTATTATTG	TTCTTTCACT	TATTCAGAAC	2195
ATTACATGCC	TTCAAAATGG	GATTGTACTA	TACCAGTAAG	TGCCACTTCT	GTGTCTTTCT	2255
AATGGAAATG	AGTAGAATTG	CTGAAAGTCT	CTATGTTAAA	ACCTATAGTG	TTT	2308

(2) INFORMATION FOR SEQ ID NO: 10: SEQUENCE CHARACTERISTICS:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 503 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

Met Glu Ala Ala Val Ala Ala Pro Arg Pro Arg Leu Leu Leu Val

1 1 5 10 15 1 5 10 15

🖳 Leu Ala Ala Ala Ala Ala Ala Ala Ala Leu Leu Pro Gly Ala Thr 20

Ala Leu Gln Cys Phe Cys His Leu Cys Thr Lys Asp Asn Phe Thr Cys 40

Val Thr Asp Gly Leu Cys Phe Val Ser Val Thr Glu Thr Thr Asp Lys 55

Val Ile His Asn Ser Met Cys Ile Ala Glu Ile Asp Leu Ile Pro Arg 65 70

Asp Arg Pro Phe Val Cys Ala Pro Ser Ser Lys Thr Gly Ser Val Thr 85 90

Thr Thr Tyr Cys Cys Asn Gln Asp His Cys Asn Lys Ile Glu Leu Pro 105

Thr Thr Val Lys Ser Ser Pro Gly Leu Gly Pro Val Glu Leu Ala Ala 115

Val Ile Ala Gly Pro Val Cys Phe Val Cys Ile Ser Leu Met Leu Met Val Tyr Ile Cys His Asn Arg Thr Val Ile His His Arg Val Pro Asn Glu Glu Asp Pro Ser Leu Asp Arg Pro Phe Ile Ser Glu Gly Thr Thr Leu Lys Asp Leu Ile Tyr Asp Met Thr Thr Ser Gly Ser Gly Ser Gly Leu Pro Leu Leu Val Gln Arg Thr Ile Ala Arg Thr Ile Val Leu Gln Glu Ser Ile Gly Lys Gly Arg Phe Gly Glu Val Trp Arg Gly Lys Trp Arg Gly Glu Glu Val Ala Val Lys Ile Phe Ser Ser Arg Glu Glu Arg Ser Trp Phe Arg Glu Ala Glu Ile Tyr Gln Thr Val Met Leu Arg His 🗎 Glu Asn Ile Leu Gly Phe Ile Ala Ala Asp Asn Lys Asp Asn Gly Thr Trp Thr Gln Leu Trp Leu Val Ser Asp Tyr His Glu His Gly Ser Leu 🛀 Phe Asp Tyr Leu Asn Arg Tyr Thr Val Thr Val Glu Gly Met Ile Lys Leu Ala Leu Ser Thr Ala Ser Gly Leu Ala His Leu His Met Glu Ile Val Gly Thr Gln Gly Lys Pro Ala Ile Ala His Arg Asp Leu Lys Ser Lys Asn Ile Leu Val Lys Lys Asn Gly Thr Cys Cys Ile Ala Asp Leu Gly Leu Ala Val Arg His Asp Ser Ala Thr Asp Thr Ile Asp Ile Ala Pro Asn His Arg Val Gly Thr Lys Arg Tyr Met Ala Pro Glu Val Leu Asp Asp Ser Ile Asn Met Lys His Phe Glu Ser Phe Lys Arg Ala Asp

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	Ile	Tyr	Ala	Met	Gly 405	Leu	Val	Phe	Trp	Glu 410	Ile	Ala	Arg	Arg	Cys 415	Ser		
	Ile	Gly	Gly	Ile 420	His	Glu	Asp	Tyr	Gln 425	Leu	Pro	Tyr	Tyr	Asp 430	Leu	Val		
	Pro	Ser	Asp 435	Pro	Ser	Val	Glu	Glu 440	Met	Arg	Lys	Val	Val 445	Cys	Glu	Gln		
	Lys	Leu 450	Arg	Pro	Asn	Ile	Pro 455	Asn	Arg	Trp	Gln	Ser 460	Cys	Glu	Ala	Leu		
	Arg 465	Val	Met	Ala	Lys	Ile 470	Met	Arg	Glu	Сув	Trp 475	Tyr	Ala	Asn	Gly	Ala 480		
	Ala	Arg	Leu	Thr	Ala 485	Leu	Arg	Ile	Lys	Lys 490	Thr	Leu	Ser	Gln	Leu 495	Ser		
L. P. L. P. U.S	Gln	Gln	Glu	Gly 500	Ile	Lys	Met											
40 1974 Hall man Hall 1981 Hall 1981 Hall 111. 1111 11	(2)	(i) (ii) (iv) (v) (vi	(A (E (I (I (I (I (A (A (E (E	EQUEN () LE () ST () ST () TC () TC () TC () CE () OF ()	ICE (PICE (PE: PE) POLO POLO POLO POLO POLO POLO POLO POLO	HARP I: 19 nucl DEDNE DEST: CAL: CAL: COM: SM:	CTEF 22 h eic ESS: line CDN NO inte E: Mous CDS 241.	RISTI pase ació unkr ar NA ernal	CS: pair l nown		 10: 1							
	GAGA	GCAC	AG C	CCTI	CCCA	G TC	:ccc	GAGO	C CGC	CGCG	CCA	CGCG	GCGCA	TG A	\TCA#	AGACCI	?	60
	TTTC	CCCG	GC C	CCAC	AGGG	C CI	CTGG	ACGI	GAG	ACCC	CGG	CCGC	CTCC	GC A	AGGA	AGAGGC	:	120
	GGGG	GTCG	AG I	CGCC	CTGT	C CA	AAGG	CCTC	: AAT	CTAA	ACA	ATCI	TGAI	TC C	TGTI	GCCGG	3	180
	CTGG	CGGG	AC C	CTGA	ATG	C AG	GAAA	TCTC	ACC	ACAT	CTC	TTCT	CCTA	TC I	CCA	AGGACC	2	240
													CTG Leu					288

				ACC Thr 20														.336
				GAG Glu														384
				GTG Val														432
				TGT Cys														480
	Thr			CTG Leu														528
1	AAC Asn	GTG Val	TCT Ser	CTG Leu 100	ATG Met	CTG Leu	GAG Glu	GCC Ala	ACC Thr 105	CAA Gln	ACT Thr	CCT Pro	TCG Ser	GAG Glu 110	GAG Glu	CCA Pro		576
<u> </u>	GAA			GCC Ala														624
				GTG Val														672
				AAG Lys													• · · · · · · · · · · · · · · · · · · ·	720
				AAG Lys														768
				GAC Asp 180						Gly								816
				ACG Thr														864
				TAT Tyr														912

							TCC Ser										96	50
	GAG					AAC	ACA Thr				AGA					CTA	100	8
							ATG Met										105	56
							CAT His										110)4
	Gln						CCC Pro 295										115	52
ئيسية 1 - 1							CAC His										120	00
	GGC Gly	AAA Lys	CCA Pro	GCC Ala	ATT Ile 325	GCC Ala	CAT His	CGT Arg	GAC Asp	CTC Leu 330	AAG Lys	AGT Ser	CGC Arg	AAT Asn	GTG Val 335	CTG Leu	124	18
							TGT Cys										129	96
							GAG Glu										134	4
							ATG Met 375	Ala									139	92
							TCG Ser										144	10
							ATC Ile										148	88
							CCT Pro										153	36

									GTG Val							CCC Pro	1584
									GAT Asp								1632
(CCC Pro							-	1680
									CAG Gln								1728
				GTG Val 500			TAGO	CCAC	GGG (CCAC	CAGGO	CT TO	CTCI	rgcci	ŗ		1776
	AAA	TGT	TG (CTGG	GAA(GA AC	ACAT	AGC	TGT	CTG	GTA	GAGO	GAGI	GA A	AGAGA	AGTGTG	1836
Ų.	CACGCTGCCC TGTGTGTGCC TGCTCAGCTT GCTCCCAGCC CATCCAGCCA AAAATACAGC											TACAGC	1896				
£ :													1922				
74											٠						
(2) INFORMATION FOR SEQ ID NO: 12: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 502 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:																	
u U		(ii	(<i>I</i> (E (I (L) MC	A) LECTOLECT	ENGTI (PE: (POL) JLE 1	H: 50 amir OGY: TYPE:	2 and a control of the control of th	nino :id :ar :teir	ació		·····		- 4			. •	
	· · · · ·	(ii (xi	() () () () M() SE	A) LEGOLECUENTE	ENGTI (PE:)POL()LE] (CE I	A: 50 amir OGY: TYPE: DESCR	2 am no ac line pro	nino id ear oteir	ació SEQ	ID N			. <u>.</u>			21.0	
	Met 1	(ii (xi	() () () () M() SE	A) LEGOLECUENTE	ENGTI (PE:)POL()LE] (CE I	A: 50 amir OGY: TYPE: DESCR	2 am no ac line pro	nino id ear oteir	ació	ID N			Leu	Ser	Val	Ala	
ŀ	1	(ii (xi	(1 (I (I) M(L) SI Leu	A) LEGOLECT	ENGTH (PE:)POLO)LE 1 NCE I Ser 5	A: 50 amir OGY: TYPE: DESCE	2 and according to the second	nino :id ear oteir ON:	ació SEQ	ID N Leu 10	Leu	Met			15		
I	1 Leu	(iii (xi Thr	(1) (I) (I) (I) (I) (I) (I) (I) (I) (I) (I	A) LE B) TY D) TO DLECT EQUEN Gly Thr 20	ENGTH YPE: DPOLO JLE T NCE I Ser 5	H: 50 amir OGY: TYPE: DESCF Phe	02 am no ac line pro RIPTI Arg	nino id ear oteir ON: Arg	ació SEQ Gly	ID N Leu 10 Lys	Leu Pro	Met Ser	Lys	Leu 30	15 Val	Asn	
1	1 Leu Cys	(ii (xi Thr Gly	(P (I) MC Leu Leu Cys 35	A) LE B) TY D) TO DLECT EQUEN Gly Thr 20	ENGTH YPE: DPOLO JLE T NCE I Ser 5 Gln	H: 50 amir OGY: TYPE: DESCR Phe Gly Pro	22 am 10 ac 1ine 1 pro RIPTI Arg Arg	nino eid ear oteir CON: Arg Leu Cys 40	ació SEQ Gly Ala 25	ID N Leu 10 Lys Arg	Leu Pro Pro	Met Ser Phe	Lys Cys 45	Leu 30 Gln	15 Val Gly	Asn	

Thr Glu Phe Leu Asn His His Cys Cys Tyr Arg Ser Phe Cys Asn His Asn Val Ser Leu Met Leu Glu Ala Thr Gln Thr Pro Ser Glu Glu Pro Glu Val Asp Ala His Leu Pro Leu Ile Leu Gly Pro Val Leu Ala Leu Pro Val Leu Val Ala Leu Gly Ala Leu Gly Leu Trp Arg Val Arg Arg Arg Gln Glu Lys Gln Arg Asp Leu His Ser Asp Leu Gly Glu Ser Ser Leu Ile Leu Lys Ala Ser Glu Gln Ala Asp Ser Met Leu Gly Asp Phe Leu Asp Ser Asp Cys Thr Thr Gly Ser Gly Ser Gly Leu Pro Phe Leu **Val Gln Arg Thr Val Ala Arg Gln Val Ala Leu Val Glu Cys Val Gly** Lys Gly Arg Tyr Gly Glu Val Trp Arg Gly Ser Trp His Gly Glu Ser __ Val Ala Val Lys Ile Phe Ser Ser Arg Asp Glu Gln Ser Trp Phe Arg □ 225 Glu Thr Glu Ile Tyr Asn Thr Val Leu Leu Arg His Asp Asn Ile Leu 🛱 Gly Phe Ile Ala Ser Asp Met Thr Ser Arg Asn Ser Ser Thr Gln Leu Trp Leu Ile Thr His Tyr His Glu His Gly Ser Leu Tyr Asp Phe Leu Gln Arg Gln Thr Leu Glu Pro Gln Leu Ala Leu Arg Leu Ala Val Ser Pro Ala Cys Gly Leu Ala His Leu His Val Glu Ile Phe Gly Thr Gln Gly Lys Pro Ala Ile Ala His Arg Asp Leu Lys Ser Arg Asn Val Leu Val Lys Ser Asn Leu Gln Cys Cys Ile Ala Asp Leu Gly Leu Ala Val

J

Met His Ser Gln Ser Asn Glu Tyr Leu Asp Ile Gly Asn Thr Pro Arg
355 360 365

Val Gly Thr Lys Arg Tyr Met Ala Pro Glu Val Leu Asp Glu His Ile

Val Gly Thr Lys Arg Tyr Met Ala Pro Glu Val Leu Asp Glu His Ile 370 375 380

Arg Thr Asp Cys Phe Glu Ser Tyr Lys Trp Thr Asp Ile Trp Ala Phe 385 390 395 400

Gly Leu Val Leu Trp Glu Ile Ala Arg Arg Thr Ile Ile Asn Gly Ile 405 410 415

Val Glu Asp Tyr Arg Pro Pro Phe Tyr Asp Met Val Pro Asn Asp Pro 420 425 430

Ser Phe Glu Asp Met Lys Lys Val Val Cys Val Asp Gln Gln Thr Pro 435 440 445

Thr Ile Pro Asn Arg Leu Ala Ala Asp Pro Val Leu Ser Gly Leu Ala 455 460

Gln Met Met Arg Glu Cys Trp Tyr Pro Asn Pro Ser Ala Arg Leu Thr 465 470 475 480

Ala Leu Arg Ile Lys Lys Thr Leu Gln Lys Leu Ser His Asn Pro Glu
495
495

Lys Pro Lys Val Ile His 500

- (2) INFORMATION FOR SEQ ID NO: 13:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2070 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (v) FRAGMENT TYPE: internal
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Mouse
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 217..1812
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

TAGO	CACA	TC I	CTGA	GAAI	T CI	'GAAG	AAAG	CAG	CAGG	STGA	AAGI	CATI	rgc c	'AAGT	'GATTT	120
TGT	CTGI	'AA G	GAAG	CCTC	C CI	CATI	CACI	TAC	ACCA	GTG	AGAC	CAGCA	GG A	CCAG	TCATT	180
CAAI	AGGGC	CG I	'GTAC	CAGGA	C GC	GTGG	CAAT	CAG	ACA	ATG Met 1	ACT Thr	CAG Gln	CTA Leu	TAC Tyr	ACT Thr	234
TAC Tyr	ATC Ile	AGA Arg	TTA Leu 10	CTG Leu	GGA Gly	GCC Ala	TGT Cys	CTG Leu 15	TTC Phe	ATC Ile	ATT Ile	TCT Ser	CAT His 20	GTT Val	CAA Gln	282
GGG Gly	CAG Gln	AAT Asn 25	CTA Leu	GAT Asp	AGT Ser	ATG Met	CTC Leu 30	CAT His	GGC Gly	ACT Thr	GGT Gly	ATG Met 35	AAA Lys	TCA Ser	GAC Asp	330
TTG Leu	GAC Asp 40	CAG Gln	AAG Lys	AAG Lys	CCA Pro	GAA Glu 45	AAT Asn	GGA Gly	GTG Val	ACT Thr	TTA Leu 50	GCA Ala	CCA Pro	GAG Glu	GAT Asp	378
□ ACC □ Thr □ 55	TTG Leu	CCT Pro	TTC Phe	TTA Leu	AAG Lys 60	TGC Cys	TAT Tyr	TGC Cys	TCA Ser	GGA Gly 65	CAC His	TGC Cys	CCA Pro	GAT Asp	GAT Asp 70	426
GCT Ala	ATT Ile	AAT Asn	AAC Asn	ACA Thr 75	TGC Cys	ATA Ile	ACT Thr	AAT Asn	GGC Gly 80	CAT His	TGC Cys	TTT Phe	GCC Ala	ATT Ile 85	ATA Ile	474
□ GAA U Glu	GAA Glu	GAT Asp	GAT Asp 90	CAG Gln	GGA Gly	GAA Glu	ACC Thr	ACA Thr 95	TTA Leu	ACT Thr	TCT Ser	GGG Gly	TGT Cys 100	ATG Met	AAG Lys	522
TAT Tyr	GAA Glu	GGC Gly 105	TCT Ser	GAT Asp	TTT Phe	CAA Gln	TGC Cys 110	AAG Lys	GAT Asp	TCA Ser	CCG Pro	AAA Lys 115	GCC Ala	CAG Gln	CTA Leu	570
CGC Arg	AGG Arg 120	ACA Thr	ATA Ile	GAA Glu	TGT Cys	TGT Cys 125	CGG Arg	ACC Thr	AAT Asn	TTG Leu	TGC Cys 130	AAC Asn	CAG Gln	TAT Tyr	TTG Leu	618
CAG Gln 135	CCT Pro	ACA Thr	CTG Leu	CCC Pro	CCT Pro 140	GTT Val	GTT Val	ATA Ile	GGT Gly	CCG Pro 145	TTC Phe	TTT Phe	GAT Asp	GGC Gly	AGC Ser 150	666
ATC Ile	CG A Arg	TGG Trp	CTG Leu	GTT Val 155	GTG Val	CTC Leu	ATT Ile	TCC Ser	ATG Met 160	GCT Ala	GTC Val	TGT Cys	ATA Ile	GTT Val 165	GCT Ala	714
ATG Met	ATC Ile	ATC Ile	TTC Phe 170	TCC Ser	AGC Ser	TGC Cys	TTT Phe	TGC Cys 175	Tyr	AAG Lys	CAT His	TAT Tyr	TGT Cys 180	AAG Lys	AGT Ser	762

2	ATC Ile	TCA Ser	AGC Ser 185	AGG Arg	GGT Gly	CGT Arg	TAC Tyr	AAC Asn 190	CGT Arg	GAT Asp	TTG Leu	GAA Glu	CAG Gln 195	GAT Asp	GAA Glu	GCA Ala	810
			CCA						AAA Lys								858
:	AGC Ser 215	TCT Ser	GGG Gly	AGT Ser	GGA Gly	TCT Ser 220	GGA Gly	TTG Leu	CCT Pro	TTA Leu	TTG Leu 225	GTT Val	CAG Gln	CGA Arg	ACT Thr	ATT Ile 230	906
(GCC Ala	AAA Lys	CAG Gln	ATT Ile	CAG Gln 235	ATG Met	GTT Val	CGG Arg	CAG Gln	GTT Val 240	GGT Gly	AAA Lys	GGC Gly	CGC Arg	TAT Tyr 245	GGA Gly	954
(GAA Glu	GTA Val	TGG Trp	ATG Met 250	GGT Gly	AAA Lys	TGG Trp	CGT Arg	GGT Gly 255	GAA Glu	AAA Lys	GTG Val	GCT Ala	GTC Val 260	AAA Lys	GTG Val	1002
	TTT Phe	TTT Phe	ACC Thr 265	ACT Thr	GAA Glu	GAA Glu	GCT Ala	AGC Ser 270	TGG Trp	TTT Phe	AGA Arg	GAA Glu	ACA Thr 275	GAA Glu	ATC Ile	TAC Tyr	1050
	CAG Gln	ACG Thr 280	GTG Val	TTA Leu	ATG Met	CGT Arg	CAT His 285	GAA Glu	AAT Asn	ATA Ile	CTT Leu	GGT Gly 290	TTT Phe	ATA Ile	GCT Ala	GCA Ala	1098
	GAC Asp 295	ATT Ile	AAA Lys	GGC Gly	ACT Thr	GGT Gly 300	TCC Ser	TGG Trp	ACT Thr	CAG Gln	CTG Leu 305	TAT Tyr	TTG Leu	ATT Ile	ACT Thr	GAT Asp 310	1146
Ų,	TAC								GAC Asp								1194
	GAC Asp	ACC Thr	AGA Arg	GCC Ala 330	CTA Leu	CTC Leu	AAG Lys	TTA Leu	GCT Ala 335	TAT Tyr	TCT Ser	GCT Ala	GCT Ala	TGT Cys 340	GGT Gly	CTG Leu	1242
									GGT Gly								1290
									AAC Asn								1338
	AGT Ser 375	Cys	TGT Cys	ATT Ile	GCT Ala	GAC Asp 380	CTG Leu	GGC Gly	CTA Leu	GCT Ala	GTT Val 385	AAA Lys	TTC Phe	AAC Asn	AGT Ser	GAT Asp 390	1386

							CCC Pro										1434
							CTG Leu										1482
							GAC Asp										1530
							ATT Ile 445										1578
							GTG Val										1626
==							AAA Lys										1674
							CTT Leu									GAA Glu	1722
- : ==:							GCC Ala										1770
Ļ	AAG Lys	ACA Thr 520	CTT Leu	GCA Ala	AAA Lys	ATG Met	GTT Val 525	GAA Glu	TCC Ser	CAG Gln	GAT Asp	GTA Val 530	AAG Lys	ATT Ile			1812
	TGAC	'AAT'I	'AA A	CAAI	TTTT	A GO	GAG	ATTI	r AGA	ACTGO	CAAG	AACT	TCT	rca (CCCA	AGGAAT	1872
	GGGI	GGG	ATT 2	AGCAI	rggaz	AT AC	GAT	TTG#	A CT	rggti	TCC	AGAC	CTCC	TTC (CTCT	CATCT	1932
	TCAC	AGGC	TG (CTAAC	CAGT	AA AC	CTT	ACCGI	r act	CTAC	CAGA	ATA	CAAG	ATT (GAA (CTTGGA	1992
	ACTI	CAAA	CA 1	GTC	ATTCI	T T	TAT	ATGAC	C AGO	CTTTC	TTT	TAAT	rgrgo	GG 1	CTTT	TTGTT	2052
	TGCI	TTTT	TT C	TTTT	rgtt												2070

- (2) INFORMATION FOR SEQ ID NO: 14:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 532 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

Met Thr Gln Leu Tyr Thr Tyr Ile Arg Leu Leu Gly Ala Cys Leu Phe 1 5 10 15

Ile Ile Ser His Val Gln Gly Gln Asn Leu Asp Ser Met Leu His Gly 20 25 . 30

Thr Gly Met Lys Ser Asp Leu Asp Gln Lys Lys Pro Glu Asn Gly Val
35 40 45

Thr Leu Ala Pro Glu Asp Thr Leu Pro Phe Leu Lys Cys Tyr Cys Ser 50 55 60

④Gly His Cys Pro Asp Asp Ala Ile Asn Asn Thr Cys Ile Thr Asn Gly □ 65 70 75 80

His Cys Phe Ala Ile Ile Glu Glu Asp Asp Gln Gly Glu Thr Thr Leu

85 90 95

Thr Ser Gly Cys Met Lys Tyr Glu Gly Ser Asp Phe Gln Cys Lys Asp

Ser Pro Lys Ala Gln Leu Arg Arg Thr Ile Glu Cys Cys Arg Thr Asn 115 120 125

Leu Cys Asn Gln Tyr Leu Gln Pro Thr Leu Pro Pro Val Val Ile Gly 130 135 140

Pro Phe Phe Asp Gly Ser Ile Arg Trp Leu Val Val Leu Ile Ser Met 145 150 155 160

Ala Val Cys Ile Val Ala Met Ile Ile Phe Ser Ser Cys Phe Cys Tyr 165 170 175

Lys His Tyr Cys Lys Ser Ile Ser Ser Arg Gly Arg Tyr Asn Arg Asp 180 185 190

Leu Glu Gln Asp Glu Ala Phe Ile Pro Val Gly Glu Ser Leu Lys Asp 195 200 205

Leu Ile Asp Gln Ser Gln Ser Ser Gly Ser Gly Ser Gly Leu Pro Leu 210 215 220

Leu Val Gln Arg Thr Ile Ala Lys Gln Ile Gln Met Val Arg Gln Val Gly Lys Gly Arg Tyr Gly Glu Val Trp Met Gly Lys Trp Arg Gly Glu Lys Val Ala Val Lys Val Phe Phe Thr Thr Glu Glu Ala Ser Trp Phe Arg Glu Thr Glu Ile Tyr Gln Thr Val Leu Met Arg His Glu Asn Ile Leu Gly Phe Ile Ala Ala Asp Ile Lys Gly Thr Gly Ser Trp Thr Gln Leu Tyr Leu Ile Thr Asp Tyr His Glu Asn Gly Ser Leu Tyr Asp Phe Leu Lys Cys Ala Thr Leu Asp Thr Arg Ala Leu Leu Lys Leu Ala Tyr Ser Ala Ala Cys Gly Leu Cys His Leu His Thr Glu Ile Tyr Gly Thr 🗎 Gln Gly Lys Pro Ala Ile Ala His Arg Asp Leu Lys Ser Lys Asn Ile E Leu Ile Lys Lys Asn Gly Ser Cys Cys Ile Ala Asp Leu Gly Leu Ala Ual Lys Phe Asn Ser Asp Thr Asn Glu Val Asp Ile Pro Leu Asn Thr Arg Val Gly Thr Lys Arg Tyr Met Ala Pro Glu Val Leu Asp Glu Ser Leu Asn Lys Asn His Phe Gln Pro Tyr Ile Met Ala Asp Ile Tyr Ser Phe Gly Leu Ile Ile Trp Glu Met Ala Arg Arg Cys Ile Thr Gly Gly Ile Val Glu Glu Tyr Gln Leu Pro Tyr Tyr Asn Met Val Pro Ser Asp Pro Ser Tyr Glu Asp Met Arg Glu Val Val Cys Val Lys Arg Leu Arg Pro Ile Val Ser Asn Arg Trp Asn Ser Asp Glu Cys Leu Arg Ala Val

Leu Lys Leu Met Ser Glu Cys Trp Ala His Asn Pro Ala Ser Arg Leu

Thr	Ala	Leu 515	Arg	Ile	Lys	Lys	Thr 520	Leu	Ala	Lys	Met	Val 525	Glu	Ser	Gln	
Asp	Val 530	Lys	Ile													
(2)	INI) SI (1 (1	EQUEI A) LI B) TI C) SI	NCE (ENGTI YPE: TRANI	R SE(CHARA H: 2] nucl DEDNI	ACTEI L60 l Leic ESS:	RIST: pase acio unki	ICS: pai: i	cs			÷				50 144
	(i: (i: (v) (v:	i) M(ii) I 7) Al 7) FRI i) Ol (1	OLECT HYPO: NTI - S AGMEN RIGIN A) ON	ULE THETE SENSI NT TENAL SE	TYPE: ICAL: E: NO YPE: SOURO ISM:	: cD1 : NO) inte	NA erna:	l							٠.	-
	·	(1	3) L(AME/I	KEY: ION: DESCI	10.			ID 1	10: :	15:					
CGC(GTT <i>l</i>				AG TO lu Se						er Pl					48
	GTC	Me CTC	t Al 1 CTG	la G] CTC	lu Se GCC	GGC	La G: 5 AGC	ly Al	La Se GGG	er Se	er Pl	ne Pl LO CCC	ne Pr CGG	co Le	eu ATC	96
GTT Val	GTC Val 15 GCT	CTC Leu CTG	CTG Leu	CTC Leu TGT	lu Se GCC	GGC Gly 20	La G 5 AGC Ser ACC	GGC Gly AGC	GGG Gly TGC	TCC Ser	GGG Gly 25 CAG	CCC Pro	CGG Arg	GGG Gly TAC	ATC Ile	
GTT Val CAG Gln 30	GTC Val 15 GCT Ala	CTC Leu CTG Leu	CTG Leu CTG Leu	CTC Leu TGT Cys	GCC Ala GCG Ala	GGC Gly 20 TGC Cys	AGC Ser ACC Thr	GGC Gly AGC Ser	GGG Gly TGC Cys	TCC Ser CTA Leu 40	GGG Gly 25 CAG Gln	CCC Pro ACC Thr	CGG Arg AAC ASn	GGG Gly TAC Tyr	ATC Ile ACC Thr 45	96
GTT Val CAG Gln 30 TGT Cys	GTC Val 15 GCT Ala GAG Glu	CTC Leu CTG Leu ACA Thr	CTG Leu CTG Leu CAT Asp	CTC Leu TGT Cys GGG Gly 50	GCC Ala GCG Ala 35	GGC Gly 20 TGC Cys	AGC Ser ACC Thr ATG Met	GGC Gly AGC Ser GTC Val	GGG Gly TGC Cys TCC Ser 55	TCC Ser CTA Leu 40 ATC	GGG Gly 25 CAG Gln TTT Phe	CCC Pro ACC Thr AAC Asn	CGG Arg AAC Asn CTG Leu	GGG Gly TAC Tyr GAT Asp 60	ATC Ile ACC Thr 45 GGC Gly	96

	TGC Cys 95																336
	GGA Gly																384
	CTG Leu																432
	ATC Ile															سر معطيد المعادد	480
	CGC Arg																528
TCC USer U																	576
GGG Gly 190	Ser	Gly	Ser	Gly	Leu 195	Pro	Leu	Phe	Val	Gln 200	Arg	Thr	Val	Ala	Arg 205		624
	TIE	vaı	ьеu	210	GIU	TIE	TIE	GIĀ	Lув 215	GIĀ	Arg	Pne	GIA	220	Val		672
∰ TGG ∭ Trp	CGT Arg	GGT Gly	CGC Arg 225	TGG Trp	AGG Arg	GGT Gly	GGT Gly	GAC Asp 230	GTG Val	GCT Ala	GTG Val	AAA Lys	ATC Ile 235	TTC Phe	TCT Ser		720
	CGT Arg																768
	ATG Met 255																816
	GAT Asp														-		864

							GAT Asp											912
							GCC Ala											960
							GGC Gly										1	1008
							AAC Asn 340										.1	1056
							CTG Leu										1	1104
	ml						AAT Asn										:	1152
Ī.	GCT Ala	CCT Pro	GAA Glu	GTC Val 385	CTT Leu	GAC Asp	GAG Glu	ACA Thr	ATC Ile 390	AAC Asn	ATG Met	AAG Lys	CAC His	TTT Phe 395	GAC Asp	TCC Ser	:	1200
							TAT Tyr										•	1248
							GGA Gly 420										:	1296
							TCC Ser										:	1344
							CTA Leu										:	1392
							GTG Val										:	1440
							CGT Arg										:	1488

		Gln	CTA Leu										GCT6	TTC			1534
CTC	TGCC	TAC	ACAA.	AGAA	CC T	GGGC	agtg:	A GG	ATGA	CTGC	AGC	CACC	GTG	CAAG	CGTC	3T	1594
GGA	.GGCC	TAT	CCTC	TTGT'	TT C	TGCC	CGGC	С СТ	CTGG	CAGA	GCC	CTGG	CCT	GCAA	GAGG	3A	1654
CAG	AGCC	TGG	GAGA	CGCG	CG C	ACTC	CCGT'	r gg	GTTT	GAGA	CAG	ACAC	TTT	TTAT	ATTT?	AC	1714
CTC	CTGA	TGG	CATG	GAGA	CC T	GAGC	AAAT(C AT	GTAG	TCAC	TCA	ATGC	CAC	AACT	CAAAC	CT	1774
GCT	TCAG	TGG	GAAG'	raca(GA G	ACCC	AGTG	CAT	TGCG	TGTG	CAG	GAGC	GTG	AGGT	GCTGG	G	1834
CTC	GCCA	GGA	GCGG	cccc	CA T	ACCT:	rgtgo	3 TC	CACT	GGGC	TGC	AGGT	TTT	CCTC	CAGGG	BA	1894
CCA	GTCA	ACT	GGCA:	rcaa(GA T	ATTG/	AGAG	AA E	CCGG.	AAGT	TTC	rccc	TCC	TTCC	CGTAG	€C	1954
AGT	CCTG	AGC	CACA	CCAT	CC T	rctc2	ATGG!	A CA	TCCG	GAGG	ACT	GCCC	CTA	GAGA	CACAA	/C	2014
<u>C</u> CTG	CTGC	CTG	TCTG:	CCAC	GC C	AAGT	GCGC2	A TG	TGCC	GAGG	TGT	3TCC	CAC	ATTG'	IGCCI	:G	2074
jgtc D	TGTG	CCA	CGCC	CGTGI	rg T	GTGT	STGT	G TG	TGTG	AGTG	AGT	STGT	GTG	TGTA	CACTI	'A	2134
-ACC	TGCT'	TGA	GCTT	CTGT	GC A	rgtgi	ľ							•			2160
뒢																	
(2) (3) (3) (3) (4) (5) (4) (5) (6) (6) (6) (6) (6) (6) (6) (6) (6) (6	(i) (i:) S: (; (; i) M:	ATION EQUEN A) LI B) TY D) TO OLECU EQUEN	NCE C ENGTH PE: POLC JLE T	HARA I: 50 amir GY: YPE:	ACTER 05 and no acc line pro	RISTI mino cid ear oteir	CS: acio		NO: 1	.6 :		. .	·		71	
Met 1	Ala	Glu	Ser	Ala 5	Gly	Ala	Ser	Ser	Phe 10	Phe	Pro	Leu	Val	Val 15	Leu		
Leu	Leu	Ala	Gly 20	Ser	Gly	Gly	Ser	Gly 25	Pro	Arg	Gly	Ile	Gln 30	Ala	Leu		
Leu	Cys	Ala 35	Cys	Thr	Ser	Cys	Leu 40	Gln	Thr	Asn	Tyr	Thr 45	Cys	Glu	Thr		
Asp	Gly 50	Ala	Cys	Met	Val	Ser 55	Ile	Phe	Asn	Leu	Asp 60	Gly	Val	Glu	His		
His 65	Val	Arg	Thr	Cys	Ile 70	Pro	Lys	Val	Glu	Leu 75	Val	Pro	Ala	Gly	Lys 80		

Pro Phe Tyr Cys Leu Ser Ser Glu Asp Leu Arg Asn Thr His Cys Cys 90 Tyr Ile Asp Phe Cys Asn Lys Ile Asp Leu Arg Val Pro Ser Gly His Leu Lys Glu Pro Ala His Pro Ser Met Trp Gly Pro Val Glu Leu Val 120 Gly Ile Ile Ala Gly Pro Val Phe Leu Leu Phe Leu Ile Ile Ile 135 130 Val Phe Leu Val Ile Asn Tyr His Gln Arg Val Tyr His Asn Arg Gln 145 Arg Leu Asp Met Glu Asp Pro Ser Cys Glu Met Cys Leu Ser Lys Asp 170 Lys Thr Leu Gln Asp Leu Val Tyr Asp Leu Ser Thr Ser Gly Ser Gly 185 180 Ser Gly Leu Pro Leu Phe Val Gln Arg Thr Val Ala Arg Thr Ile Val 200 195 W Leu Gln Glu Ile Ile Gly Lys Gly Arg Phe Gly Glu Val Trp Arg Gly Arg Trp Arg Gly Gly Asp Val Ala Val Lys Ile Phe Ser Ser Arg Glu 240 Glu Arg Ser Trp Phe Arg Glu Ala Glu Ile Tyr Gln Thr Val Met Leu 255 245 u 🖺 Arg His Glu Asn Ile Leu Gly Phe Ile Ala Ala Asp Asn Lys Asp Asn 260 265 Gly Thr Trp Thr Gln Leu Trp Leu Val Ser Asp Tyr His Glu His Gly 280 Ser Leu Phe Asp Tyr Leu Asn Arg Tyr Thr Val Thr Ile Glu Gly Met 300 295 290 Ile Lys Leu Ala Leu Ser Ala Ala Ser Gly Leu Ala His Leu His Met 305 310 Glu Ile Val Gly Thr Gln Gly Lys Pro Gly Ile Ala His Arg Asp Leu Lys Ser Lys Asn Ile Leu Val Lys Lys Asn Gly Met Cys Ala Ile Ala 345

Asp Leu Gly Leu Ala Val Arg His Asp Ala Val Thr Asp Thr Ile Asp 365 360 355 Ile Ala Pro Asn Gln Arg Val Gly Thr Lys Arg Tyr Met Ala Pro Glu 375 370 Val Leu Asp Glu Thr Ile Asn Met Lys His Phe Asp Ser Phe Lys Cys 385 Ala Asp Ile Tyr Ala Leu Gly Leu Val Tyr Trp Glu Ile Ala Arg Arg 405 Cys Asn Ser Gly Gly Val His Glu Asp Tyr Gln Leu Pro Tyr Tyr Asp 420 Leu Val Pro Ser Asp Pro Ser Ile Glu Glu Met Arg Lys Val Val Cys 445 440 435

Asp Gln Lys Leu Arg Pro Asn Val Pro Asn Trp Trp Gln Ser Tyr Glu 450 455 460

Ala Leu Arg Val Met Gly Lys Met Met Arg Glu Cys Trp Tyr Ala Asn
470 475 480

Gly Ala Ala Arg Leu Thr Ala Leu Arg Ile Lys Lys Thr Leu Ser Gln
485 490 495

Leu Ser Val Gln Glu Asp Val Lys Ile
500 505

- (2) INFORMATION FOR SEQ ID NO: 17:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1952 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown
 - (ii) MOLECULE TYPE: cDNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (v) FRAGMENT TYPE: internal
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Mouse
 - (ix) FEATURE:

<u>_</u>

- (A) NAME/KEY: CDS
- (B) LOCATION: 187..1692
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

TGGGAAGCGG CGGCGGTTA ACTTCGGCTG AATCACAACC ATTTGGCGCT GAGCTATG	AC 120
AAGAGAGCAA ACAAAAAGTT AAAGGAGCAA CCCGGCCATA AGTGAAGAGA GAAGTTTA	TT 180
GATAAC ATG CTC TTA CGA AGC TCT GGA AAA TTA AAT GTG GGC ACC AAG Met Leu Leu Arg Ser Ser Gly Lys Leu Asn Val Gly Thr Lys 1 5 10	228
AAG GAG GAT GGA GAG AGT ACA GCC CCC ACC CCT CGG CCC AAG ATC CTA Lys Glu Asp Gly Glu Ser Thr Ala Pro Thr Pro Arg Pro Lys Ile Leu 15 20 25 30	
CGT TGT AAA TGC CAC CAC CAC TGT CCG GAA GAC TCA GTC AAC AAT ATC Arg Cys Lys Cys His His His Cys Pro Glu Asp Ser Val Asn Asn Ile 35 40 45	324
TGC AGC ACA GAT GGG TAC TGC TTC ACG ATG ATA GAA GAA GAT GAC TCT Cys Ser Thr Asp Gly Tyr Cys Phe Thr Met Ile Glu Glu Asp Asp Ser 50 55 60	
GGA ATG CCT GTT GTC ACC TCT GGA TGT CTA GGA CTA GAA GGG TCA GAT Gly Met Pro Val Val Thr Ser Gly Cys Leu Gly Leu Glu Gly Ser Asp 65 70 75	420
TTT CAA TGT CGT GAC ACT CCC ATT CCT CAT CAA AGA AGA TCA ATT GAA Phe Gln Cys Arg Asp Thr Pro Ile Pro His Gln Arg Arg Ser Ile Glu 80 85 90	468
TGC TGC ACA GAA AGG AAT GAG TGT AAT AAA GAC CTC CAC CCC ACT CTG UCys Cys Thr Glu Arg Asn Glu Cys Asn Lys Asp Leu His Pro Thr Leu 95 100 105 110	
© CCT CCT CTC AAG GAC AGA GAT TTT GTT GAT GGG CCC ATA CAC CAC AAG © Pro Pro Leu Lys Asp Arg Asp Phe Val Asp Gly Pro Ile His His Lys 115 120 125	
GCC TTG CTT ATC TCT GTG ACT GTC TGT AGT TTA CTC TTG GTC CTC ATT Ala Leu Leu Ile Ser Val Thr Val Cys Ser Leu Leu Leu Val Leu Ile 130 135 140	612
ATT TTA TTC TGT TAC TTC AGG TAT AAA AGA CAA GAA GCC CGA CCT CGG Ile Leu Phe Cys Tyr Phe Arg Tyr Lys Arg Gln Glu Ala Arg Pro Arg 145 150 155	
TAC AGC ATT GGG CTG GAG CAG GAC GAG ACA TAC ATT CCT CCT GGA GAG Tyr Ser Ile Gly Leu Glu Gln Asp Glu Thr Tyr Ile Pro Pro Gly Glu 160 165 170	
TCC CTG AGA GAC TTG ATC GAG CAG TCT CAG AGC TCG GGA AGT GGA TCA Ser Leu Arg Asp Leu Ile Glu Gln Ser Gln Ser Ser Gly Ser Gly Ser 175 180 185 190	

			CTG Leu													804
			ATT Ile 210													852
			GAA Glu													900
			TTC Phe													948
			ATT Ile													996
TCC Ser																1044
CTT																1092
AAG Lys																1140
WATC Wile																1188
			ATC Ile													1236
			GCT Ala													1284
CCA Pro	CCC Pro	AAC Asn	ACC Thr 370	CGG Arg	GTT Val	GGC Gly	ACC Thr	AAG Lys 375	CGC Arg	TAT Tyr	ATG Met	CCT Pro	CCA Pro 380	GAA Glu	GTG Val	1332
			AGC Ser													1380

														AGA Arg		1428
														GAC Asp		1476
														TGC Cys 445		1524
														GAG Glu		1572
														AAT Asn		1620
GCC Ala	TCC Ser 480	AGG Arg	CTG Leu	ACG Thr	GCC Ala	CTG Leu 485	AGA Arg	GTT Val	AAG Lys	AAA Lys	ACC Thr 490	CTT Leu	GCC Ala	AAA Lys	ATG Met	1668
TCA Ser 495								TGAC	CGTC	AGA I	TACTI	GTG	GA CA	AGAGO	CAAGA	1722
ATTI	CACA	AGA A	AGCAI	CGTI	'A GO	CCAA	\GCC1	TGA	ACGI	TAG	CCTA	CTGC	CCC I	AGTG	GTTCA	1782
₩ ≟GAC I	TTCC	TG G	SAAGI	GAG	CA CO	GTGG	GCAG	ACA	CAG	AGGA	ACC	CAGA	AAC A	ACGG	TTCAT	1842
₩ ©CATO	GCTI	TC 1	GAGG	BAGG	AG AA	ACTO	TTTC	GGI	'AAC'I	TGT	TCA	GAT	ATG A	ATGC <i>I</i>	TGTTG	1902
CTTI	CTAA	AGA A	AGC	CTGI	TT AT	TTGA	ATT	A CCA	TTTT	TTT	ATA	AAAA	AAA			1952

(2) INFORMATION FOR SEQ ID NO: 18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 502 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

Met Leu Leu Arg Ser Ser Gly Lys Leu Asn Val Gly Thr Lys Lys Glu

1 10 15

Asp Gly Glu Ser Thr Ala Pro Thr Pro Arg Pro Lys Ile Leu Arg Cys 20 25 30

Lys Cys His His Cys Pro Glu Asp Ser Val Asn Asn Ile Cys Ser Thr Asp Gly Tyr Cys Phe Thr Met Ile Glu Glu Asp Asp Ser Gly Met Pro Val Val Thr Ser Gly Cys Leu Gly Leu Glu Gly Ser Asp Phe Gln 65 Cys Arg Asp Thr Pro Ile Pro His Gln Arg Arg Ser Ile Glu Cys Cys Thr Glu Arg Asn Glu Cys Asn Lys Asp Leu His Pro Thr Leu Pro Pro 105 Leu Lys Asp Arg Asp Phe Val Asp Gly Pro Ile His His Lys Ala Leu 120 125 Leu Ile Ser Val Thr Val Cys Ser Leu Leu Leu Val Leu Ile Ile Leu 135 130 phe Cys Tyr Phe Arg Tyr Lys Arg Gln Glu Ala Arg Pro Arg Tyr Ser **145** Ile Gly Leu Glu Gln Asp Glu Thr Tyr Ile Pro Pro Gly Glu Ser Leu Arg Asp Leu Ile Glu Gln Ser Gln Ser Ser Gly Ser Gly Leu 180 185 Pro Leu Leu Val Gln Arg Thr Ile Ala Lys Gln Ile Gln Met Val Lys 195 200 igoplus Gln Ile Gly Lys Gly Arg Tyr Gly Glu Val Trp Met Gly Lys Trp Arg Gly Glu Lys Val Ala Val Lys Val Phe Phe Thr Thr Glu Glu Ala Ser 225 230 235 240 Trp Phe Arg Glu Thr Glu Ile Tyr Gln Thr Val Leu Met Arg His Glu 245 Asn Ile Leu Gly Phe Ile Ala Ala Asp Ile Lys Gly Thr Gly Ser Trp 265 Thr Gln Leu Tyr Leu Ile Thr Asp Tyr His Glu Asn Gly Ser Leu Tyr 280 285 Asp Tyr Leu Lys Ser Thr Thr Leu Asp Ala Lys Ser Met Leu Lys Leu 290 295

Ala Tyr Ser Ser Val Ser Gly Leu Cys His Leu His Thr Glu Ile Phe Ser Thr Gln Gly Lys Pro Ala Ile Ala His Arg Asp Leu Lys Ser Lys Asn Ile Leu Val Lys Lys Asn Gly Thr Cys Cys Ile Ala Asp Leu Gly 340 Leu Ala Val Lys Phe Ile Ser Asp Thr Asn Glu Val Asp Ile Pro Pro 360 365 Asn Thr Arg Val Gly Thr Lys Arg Tyr Met Pro Pro Glu Val Leu Asp Glu Ser Leu Asn Arg Asn His Phe Gln Ser Tyr Ile Met Ala Asp Met 390 395 385 400 Tyr Ser Phe Gly Leu Ile Leu Trp Glu Ile Ala Arg Arg Cys Val Ser 410 405 415 Gly Gly Ile Val Glu Glu Tyr Gln Leu Pro Tyr His Asp Leu Val Pro √Ser Asp Pro Ser Tyr Glu Asp Met Arg Glu Ile Val Cys Met Lys Lys 435 440 Theu Arg Pro Ser Phe Pro Asn Arg Trp Ser Ser Asp Glu Cys Leu Arg 455 450 U Gln Met Gly Lys Leu Met Thr Glu Cys Trp Ala Gln Asn Pro Ala Ser 470 475 Arg Leu Thr Ala Leu Arg Val Lys Lys Thr Leu Ala Lys Met Ser Glu 490 Ser Gln Asp Ile Lys Leu

Ser Gin Asp Ile Lys Leu 500

(2)	<pre>INFORMATION FOR SEQ ID NO: 19: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: NO (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:</pre>	
GCGGA	GATCCTG TTGTGAAGGN AATATGTG	28
"GCGAT	<pre>INFORMATION FOR SEQ ID NO: 20: (i) SEQUENCE CHARACTERISTICS:</pre>	24
(2)	<pre>INFORMATION FOR SEQ ID NO: 21: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 26 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: NO (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21:</pre>	

GCGGATCCGC GATATATTAA AAGCAA

(2)	<pre>INFORMATION FOR SEQ ID NO: 22: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: YES (xi) SEQUENCE DESCRIPTION: SEQ ID</pre>	NO: 22:	
CGGA	ATTCTG GTGCCATATA		20
(2) 09029	INFORMATION FOR SEQ ID NO: 23: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 37 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single		
W .a	(D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO		
4	(iv) ANTI-SENSE: NO (xi) SEQUENCE DESCRIPTION: SEQ ID	NO • 23 •	
਼ [ਾ] •ੂ ਨਾਜਾਨਾ	AAGGGC ACATCAACTT CATTTGTGTC ACTGTT		37
J			5,
学(2) 型 型	<pre>INFORMATION FOR SEQ ID NO: 24: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 26 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear</pre>		
	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO		
	(iv) ANTI-SENSE: NO (xi) SEQUENCE DESCRIPTION: SEQ ID	NO: 24:	
GCGG	ATCCAC CATGGCGGAG TCGGCC	ı	26

(2)	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: NO
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 25:
AACA	ACCGGGC CGGCGATGAT
	INFORMATION FOR SEQ ID NO: 26: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 6 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: peptide (v) FRAGMENT TYPE: internal (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 26: Gly Xaa Gly Xaa Xaa Gly 1 5
յուր այլ այլ (2)	<pre>INFORMATION FOR SEQ ID NO: 27: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 6 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: peptide (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 27:</pre>
	Asp Phe Lys Ser Arg Asn

(2) INFORMATION FOR SEQ ID NO: 28:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 6 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 28:

Asp Leu Lys Ser Lys Asn

(2) INFORMATION FOR SEQ ID NO: 29:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 6 amino acids

(B) TYPE: amiho acid

(D) TOPOLOGY: \linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 29:

Gly Thr Lys Arg Tyr Met